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(54) Title: NOVEL ANTI-IGF-IR ANTIBODIES AND USES THEREOF

(57) Abstract: The present invention relates to novel antibodies capable of binding specifically to the human insulin-like growth factor I receptor (IGF-IR). The invention likewise comprises the use of these antibodies as a medicament for the prophylactic and/or therapeutic treatment of cancers overexpressing IGF-IR, stimulated either by IGF1 and/or IGF2, or any pathology connected with the overexpression of said receptor as well as in processes or kits for diagnosis of illnesses connected with the overexpression of the IGF-IR and/or the IGF-I / Insulin hybrid receptor.

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NOVEL ANTI-IGF-IR ANTIBODIES AND USES THEREOF

The present invention relates to novel antibodies capable of binding specifically to the human insulin-like growth factor I receptor (IGF-IR) and/or capable of specifically inhibiting the tyrosine kinase activity of said IGF-IR, especially monoclonal antibodies of murine, chimeric and humanized origin, as well as the amino acid and nucleic acid sequences coding for these antibodies. In a particular aspect, the invention relates to novel antibodies capable to efficiently inhibiting not only the binding of the ligand IGF1 to IGF-IR but also to efficiently inhibiting the binding of the other ligand, i.e. IGF2, to the same receptor. The invention likewise comprises the use of these antibodies as a medicament for the prophylactic and/or therapeutic treatment of cancers overexpressing IGF-IR, stimulated either by IGF1 and/or IGF2, or any pathology connected with the overexpression of said receptor as well as in processes or kits for diagnosis of illnesses connected with the overexpression of the IGF-IR and/or the IGF-I / Insulin hybrid receptor. The invention finally comprises products and/or compositions comprising such antibodies in combination with, for example, anti-EGFR antibodies and/or compounds and/or anti-cancer agents or agents conjugated with toxins and their use for the prevention and/or the treatment of certain cancers.

The insulin-like growth factor I receptor called IGF-IR is a receptor with tyrosine kinase activity having 70% homology with the insulin receptor IR. IGF-IR is a glycoprotein of molecular weight approximately 350,000. It is a hetero-tetrameric receptor of which each half -linked by disulfide bridges- is composed of an extracellular α -subunit and of a transmembrane β -subunit. IGF-IR binds IGF1 and IGF2 with a very high affinity ($K_d \approx 1$ nM) but is equally capable of binding to insulin with an affinity 100 to 1000 times less. Conversely, the IR binds insulin with a very high affinity although the IGFs only bind to the insulin receptor with a 100 times lower affinity. The tyrosine kinase domain of IGF-IR and of IR has a very high sequence homology although the zones of weaker homology respectively concern the cysteine-rich region situated on the α -subunit and the C-terminal part of the β -subunit. The sequence differences observed in the α -subunit are situated in the binding zone of the ligands and are therefore at the origin of the relative affinities of IGF-IR and of IR for the IGFs and

insulin respectively. The differences in the C-terminal part of the β -subunit result in a divergence in the signalling pathways of the two receptors; IGF-IR mediating mitogenic, differentiation and antiapoptosis effects, while the activation of the IR principally involves effects at the level of the metabolic pathways (Baserga et al.,
5 Biochim. Biophys. Acta, 1332:F105-126, 1997; Baserga R., Exp. Cell. Res., 253:1-6, 1999).

The cytoplasmic tyrosine kinase proteins are activated by the binding of the ligand to the extracellular domain of the receptor. The activation of the kinases in its turn involves the stimulation of different intra-cellular substrates, including IRS-1, IRS-
10 2, Shc and Grb 10 (Peruzzi F. et al., J. Cancer Res. Clin. Oncol., 125:166-173, 1999). The two major substrates of IGF-IR are IRS and Shc which mediate, by the activation of numerous effectors downstream, the majority of the growth and differentiation effects connected with the attachment of the IGFs to this receptor. The availability of substrates can consequently dictate the final biological effect connected with the activation of the
15 IGF-IR. When IRS-1 predominates, the cells tend to proliferate and to transform. When Shc dominates, the cells tend to differentiate (Valentinis B. et al., J. Biol. Chem. 274:12423-12430, 1999). It seems that the route principally involved for the effects of protection against apoptosis is the phosphatidyl-inositol 3-kinases (PI 3-kinases) route (Prisco M. et al., Horm. Metab. Res., 31:80-89, 1999; Peruzzi F. et al., J. Cancer Res.
20 Clin. Oncol., 125:166-173, 1999).

The role of the IGF system in carcinogenesis has become the subject of intensive research in the last ten years. This interest followed the discovery of the fact that in addition to its mitogenic and antiapoptosis properties, IGF-IR seems to be required for the establishment and the maintenance of a transformed phenotype. In fact, it has been
25 well established that an overexpression or a constitutive activation of IGF-IR leads, in a great variety of cells, to a growth of the cells independent of the support in media devoid of fetal calf serum, and to the formation of tumors in nude mice. This in itself is not a unique property since a great variety of products of overexpressed genes can transform cells, including a good number of receptors of growth factors. However, the
30 crucial discovery which has clearly demonstrated the major role played by IGF-IR in the transformation has been the demonstration that the R- cells, in which the gene coding for IGF-IR has been inactivated, are totally refractory to transformation by

different agents which are usually capable of transforming the cells, such as the E5 protein of bovine papilloma virus, an overexpression of EGFR or of PDGFR, the T antigen of SV 40, activated ras or the combination of these two last factors (Sell C. et al., Proc. Natl. Acad. Sci., USA, 90:11217-11221, 1993; Sell C. et al., Mol. Cell. Biol., 14:3604-3612, 1994; Morriane A. J., Virol., 69:5300-5303, 1995; Coppola D. et al., Mol. Cell. Biol., 14:4588-4595, 1994; DeAngelis T et al., J. Cell. Physiol., 164:214-221, 1995).

IGF-IR is expressed in a great variety of tumors and of tumor lines and the IGFs amplify the tumor growth via their attachment to IGF-IR. Other arguments in favor of the role of IGF-IR in carcinogenesis come from studies using murine monoclonal antibodies directed against the receptor or using negative dominants of IGF-IR. Actually, murine monoclonal antibodies directed against IGF-IR inhibit the proliferation of numerous cell lines in culture and the growth of tumor cells *in vivo* (Arteaga C. et al., Cancer Res., 49:6237-6241, 1989; Li et al., Biochem. Biophys. Res. Com., 196:92-98, 1993; Zia F. et al., J. Cell. Biol., 24:269-275, 1996; Scotlandi K. et al., Cancer Res., 58:4127-4131, 1998). It has likewise been shown in the works of Jiang et al. (Oncogene, 18:6071-6077, 1999) that a negative dominant of IGF-IR is capable of inhibiting tumor proliferation.

Such antibodies capable to bind specifically to the IGF-IR have been described and several patent applications have been filed. As an example, we can mentioned the patent application WO 03/059951, filed by the Applicant, wherein a monoclonal antibody able to bind to IGF-IR, called 7C10, is described. Others patent applications can be mentioned as WO 02/053596 (PFIZER INC. and ABGENIX INC.), WO 03/100008 (SCHERING CORPORATION) or WO 03/106621 (IMMUNOGEN INC.). All these applications are claiming antibodies capable to specifically bind to the IGF-IR and/or to inhibit its activity.

Although it is considered as an evident property for each of these antibodies, nothing in the specification of these applications clearly demonstrate that these antibodies are capable to inhibit efficiently the binding of both natural ligands to IGF-IR and, more particularly the binding IGF2 to IGF-IR. In vitro data such as inhibition of proliferation IGF1 and IGF2 induced were shown in these applications and demonstrated that the described antibodies are able to inhibit efficiently both IGF1 and

IGF2-induced proliferation. However, as the major part of described antibodies displayed the capacity of inducing a partial down-regulation of IGF-IR, it is not evident that the antiproliferative properties are directly linked to a displacement of both IGF1 and IGF2 from IGF-IR.

5 Actually, for example, data shown in WO 03/106621 are only dealing with IGF1 and under no circumstances with an eventual inhibition of the binding of the IGF2 to IGF-IR (see Example C, pages 33-35 and Figure 3, and Example D, pages 35-37 and Figures 4-6).

10 The same observations can be made for WO 02/053596 (see Example IV, page 78-79 and Figure 3, and Example VII, page 82 and Figure 4).

15 Amongst all the presently identified patent applications describing the discovery of monoclonal or recombinant antibodies directed against the human IGF-IR (hIGF-IR), only two of them (WO 2004/087756 and WO 2005/005635) are really showing an efficacy of the antibodies, respectively AK1a and AK18, to inhibit [¹²⁵I]IGF2 binding to hIGF-IR.

20 The experimental procedure, identical in both cases, is based on competition binding on human colorectal adenocarcinoma (HT29) intact cells. Although the described procedure is sound, no positive controls such as competition by natural hIGF-IR ligands (IGF1 and IGF2) is presented, making the data on competition binding suspicious. On the other hand, only incomplete competition was shown (maximally 80 % inhibition of [¹²⁵I]IGF2 binding for both AK1a and AK18 antibody), even at high antibody concentration, making the development of more efficacious antibodies necessary for enhanced therapeutic efficacy in humans. A last controversial point concerning the putative inhibition of IGF2-mediated responses by AK1a and AK18 is
25 the absence of examples showing an inhibition of functional IGF2-stimulated signaling mediated by hIGF-IR. Indeed, even if competition of IGF2 binding is happening, it must be associated with a concomitant inhibition of downstream hIGF-IR signalling and such evidence must be exemplified to provide mechanistic evidence for antibody functional efficacy.

30

 The object of the present invention is to be able to have available a murine monoclonal antibody, preferably a chimerized or humanized antibody, which will

recognize IGF-IR specifically with great affinity and which is also able to inhibit not only the binding of IGF1 to IGF-IR but also the binding of IGF2 to IGF-IR. This antibody will interact little or not at all with the IR. Its attachment will be able to inhibit the *in vitro* growth of cell lines overexpressing IGF-IR by interacting principally with the signal transduction pathways activated during IGF1/IGF-IR and IGF2/IGF-IR interactions. This antibody will be able to be active *in vivo* on all tumors types expressing IGF-IR including estrogen-dependent breast tumors and prostate tumors, which is not the case for the anti-IGF-IR monoclonal antibodies (written MAb or MAB) currently available. In effect, α IR3, which refers to the domain of IGF-IR, totally inhibits the growth of estrogen-dependent tumors of the breast (MCF-7) *in vitro* but is without effect on the corresponding model *in vivo* (Arteaga C. et al., J. Clin. Invest. 84:1418-1423, 1989). In the same way, the scFv-Fc fragment derived from the murine monoclonal 1H7 is only weakly active on the tumor of the breast MCF-7 and totally inactive on an androgen-independent tumor of the prostate (Li S. L. et al., Cancer Immunol. Immunother., 49:243-252, 2000).

In a surprising manner, the inventors have generated a murine monoclonal antibody described as I-3466, recognising IGF-IR and corresponding to all of the criteria stated above, that is to say to a nonrecognition of the receptor on the insulin, to an *in vitro* blockage of the IGF1 and particularly the IGF2 proliferation induced but likewise to the *in vivo* inhibition of the growth of different tumors expressing IGF-IR among which are an osteosarcoma and prostate tumors. Furthermore, it has been shown that these antibodies inhibit the IGF1- and/or IGF2-induced phosphorylation of the tyrosine of the beta chain of IGF-IR both on both MCF-7 and HT29 cells. Moreover, it has likewise been established that these antibodies cause the internalization of said receptor and its degradation contrary to what is usually observed with natural ligands which allow the rapid recycling of the receptor on the surface of the cells. It has been possible to characterize these antibodies by their peptidic and nucleic sequence, especially by the sequence of their regions determining their complementarity (CDR) for IGF-IR.

Thus, according to a first embodiment, a subject of the present invention is an isolated antibody, or one of its functional fragments, said antibody or one of its said fragments being capable of binding specifically to the human insulin-like growth factor

I receptor and/or capable of specifically inhibiting the tyrosine kinase activity of said IGF-IR receptor, characterized in that it is capable of inhibiting the natural attachment of its first ligand IGF1 with an IC_{50} of less than 0,3 nM, preferentially less than 0,03 nM and also capable of inhibiting the natural attachment of its second ligand IGF2 with an IC_{50} of less than 0,3 nM, preferentially less than 0,1 nM.

More particularly, the invention concerns an isolated antibody, or a functional immunogenic fragment thereof, having a binding affinity for the human insulin-like growth factor I receptor (IGF-IR), characterized in that upon binding said IGF-IR, it inhibits the binding of the native binding partner IGF1 to said IGF-IR with an IC_{50} of less than 0,3 nM, preferentially less than 0,03 nM and it also inhibits the binding of the native binding partner IGF2 to said IGF-IR with an IC_{50} of less than 0,3 nM, preferentially less than 0,1 nM.

In addition, the invention concerns an isolated antibody, or a functional immunogenic fragment thereof, having a tyrosine kinase of the human insulin-like growth factor I receptor (IGF-IR) inhibiting activity, characterized in that upon binding said IGF-IR, it inhibits the binding of the native binding partner IGF1 to said IGF-IR with an IC_{50} of less than 0,3 nM, preferentially less than 0,03 nM and it also inhibits the binding of the native binding partner IGF2 to said IGF-IR with with an IC_{50} of less than 0,3 nM, preferentially less than 0,1 nM.

In the present application, IC_{50} have been determined graphically as explained in example 2.

Regarding the state on the art, Roche antibody referenced 18 (WO 2004/087756 and WO 2005/005635) have an average IC_{50} for both for IGF1 and IGF2 which is about 0.3 nM (see example 6 of WO 2005/005635), i.e. more than the IC_{50} obtained for the antibody I-3466 (see example 2).

In the present description, the terms "to bind" and "to attach" have the same meaning and are inter-changeable.

According to another embodiment of the invention, the antibody is also capable to bind the IGF-I / Insulin hybrid receptor.

Actually, IGF-IR shows a high homology with the Insulin receptor (IR) which exists under two isoforms A and B.

Sequences of IR, isoforms A and B, are registered under Accession Numbers X02160 and M10051, respectively, in the NCBI Genbank. Other datas, without limitations, relating to IR are incorporated herein by references (Vinten et al., 1991, Proc. Natl. Acad. Sci. USA, 88:249-252; Belfiore et al., 2002, The Journal of Biological Chemistry, 277:39684-39695; Dumesic et al., 2004, The Journal of Endocrinology & Metabolism, 89(7):3561-3566).

The IGF-IR and IR are tetrameric glycoproteins composed of two extracellular α - and two transmembrane β -subunits linked by disulfide bonds. Each α -subunit, containing the ligand-binding site is approximately 130- to 135-kDa, whereas each β -subunit containing the tyrosine kinase domain is approximately 90- to 95- kDa. These receptors share more than 50 % overall amino acid sequence similarity and 84 % similarity in the tyrosine kinase domain. After ligand binding, phosphorylated receptors recruit and phosphorylate docking proteins, including the insulin receptor substrate-1 protein family (IRS1), Gab1 and Shc (Avruch, 1998, Mol.Cell. Biochem., 182, 31-48; Roth et al., 1988, Cold Spring Harbor Symp. Quant. Biol., 53, 537-543; White, 1998, Mol. Cell. Biochem. 182, 3-11; Laviola et al., 1997, J. Clin. Invest., 99, 830-837 ; Cheatham et al., 1995, Endocr. Rev., 16, 117-142), leading to the activation of different intracellular mediators. Although both the IR and IGF-IR similarly activate major signalling pathways, differences exist in the recruitment of certain docking proteins and intracellular mediators between both receptors (Sasaoka et al., 1996, Endocrinology 137, 4427-4434; Nakae et al., 2001, Endocr. Rev., 22, 818-835; Dupont and Le Roith, 2001, Horm. Res. 55, Suppl. 2, 22-26; Koval et al., 1998, Biochem. J., 330, 923-932). These differences are the basis for the predominant metabolic effects elicited by IR activation and the predominant mitogenic, transforming and anti-apoptotic effects elicited by IGF-IR activation (De Meyts et al., 1995, Ann. N.Y. Acad. Sci., 766, 388-401; Singh et al., 2000; Prisco et al., 1999, Horm. Metab. Res., 31, 80-89; Kido et al., 2001, J. Clin. Endocrinol. Metab., 86, 972-979). Insulin binds with high affinity to the IR (100-fold higher than to the IGF-IR), whereas insulin-like growth factors (IGF1 and IGF2) bind to the IGF-IR with 100-fold higher affinity than to the IR.

The human IR exists in two isoforms, IR-A and IR-B, generated by alternative splicing of the IR gene that either excludes or includes 12 amino acid residues encoded by a small exon (exon 11) at the carboxy-terminus of the IR α -subunit. The relative

abundance of IR isoforms is regulated by tissue specific and unknown factors (Moller et al., 1989, *Mol. Endocrinol.*, 3, 1263-1269; Mosthaf et al., 1990, *EMBO J.*, 9, 2409-2413). IR-B is the predominant IR isoform in normal adult tissues (adipose tissue, liver and muscle) that are major target tissues for the metabolic effects of insulin (Moller et al., 1989; Mosthaf et al., 1990). IR-A is the predominant isoform in fetal tissues and mediates fetal growth in response to IGF2 (Frasca et al., 1999, *Mol. Cell. Biol.*, 19, 3278-3288), as also suggested by genetic studies carried out in transgenic mice (DeChiara et al., 1990, *Nature* 345, 78-80; Louvi et al., 1997, *Dev. Biol.* 189, 33-48). Moreover, when cells transform and become malignant, dedifferentiation is often associated with an increased IR-A relative abundance (Pandini et al., 2002, *The Journal of Biological Chemistry*, Vol. 277, N° 42, pp39684-39695).

Given the high degree of homology, the insulin and IGF-I half-receptors (composed of one α - and one β -subunit) can heterodimerize, leading to the formation of insulin/IGF-I hybrid receptors (Hybrid-R) (Soos et al., 1990, *Biochem J.*, 270, 383-390; Kasuya et al., 1993, *Biochemistry* 32, 13531-13536; Seely et al., 1995, *Endocrinology* 136, 1635-1641; Bailyes et al., 1997, *Biochem J.*, 327, 209-215).

Both IR isoforms are equally able to form hybrids with IGF-IR. Hybrid-R, however, have different functional characteristics. Hybrid-RsB has reduced affinity for IGF1 and especially for IGF2. In contrast, Hybrid-RsA has a high affinity for IGF1 and bind also IGF2 and insulin at a physiological concentration range. The expression of Hybrid-RsA up-regulates the IGF system by two different mechanisms i) binding (with high affinity) and activation by both IGF1 and IGF2 (which do not occur with the Hybrid-RsB), ii) activation of the IGF-IR pathway after insulin binding. Insulin binding to Hybrid-RsA phosphorylates the IGF-IR β -subunit and activates an IGF-IR-specific substrate (CrkII) so that Hybrid-RsA shifts insulin to IGF-IR signaling (Pandini et al., 2002).

In several tissues, like liver, spleen or placenta, Hybrid-R are more represented than IGF-IR (Bailyes et al., 1997). As tumor tissues overexpress, or present an abnormal activation, both IGF-IR and IR-A (Frasca et al., 1999; Sciacca et al., 1999, *Oncogene* 18, 2471-2479; Vella et al., 2001, *Mol. Pathol.*, 54, 121-124), Hybrid-RsA may also be overexpressed in a variety of human malignancies, including thyroid and breast cancers providing a selective growth advantage to malignant cells able to respond by a type

IGF-IR signalisation following a stimulation by IGF1 and/or IGF2 but also by insulin at physiological concentrations (Baillyes et al., 1997; Pandini et al., 1999, Clin. Cancer Res. 5, 1935-1944; Belfiore et al., 1999, Biochimie (Paris) 81, 403-407; Frasca et al., 1999; Sciacca et al., 1999; Vella et al., 2001).

5 According to the invention, antibodies are also characterized in that they are capable of binding to the Hybrid-R and, if necessary, preferably moreover capable of inhibiting the natural attachment of the ligands insulin, IGF1 and/or IGF-2 of Hybrid-R and/or capable of specifically inhibiting the tyrosine kinase activity of said Hybrid-R.

 An object of the invention is an antibody, or one of its functional fragments,
10 characterized in that it is also capable of inhibiting 100 % of the IGF1 and/or IGF2 induced phosphorylation of the IGF-IR beta-chain (preferentially on HT29 cells).

 In another aspect, the antibody, or one of its functional fragments, according to the invention is characterized in that it does not present any agonistic intrinsic activity.

 The antibody, or one of its functional fragments, according to the invention, is
15 moreover characterized in that it is capable of inducing, using the same FACS analysis method:

- i) at least 30 % of the IGF-IR internalization on HT29 cells, and/or
- ii) at least 85 % of the IGF-IR internalization on MCF-7 cells.

 A characteristic of the antibody, or one of its functional fragments, according to
20 the invention, is its capacity of inducing, using the same FACS analysis method:

- i) at least 50 % of the IGF-IR degradation on HT29 cells, and/or
- ii) at least 65 % of the IGF-IR degradation on MCF-7 cells.

 In another embodiment, the antibody, or one of its functional fragments, according to the invention, is characterized in that it is able to inhibit both IGF1 and
25 IGF2-induced *in vitro* proliferation of MCF-7 cells with IC_{50s} at least equal to 1 nM and preferentially at least 0.7 and 0.5 nM respectively for IGF1 and IGF2 experiments.

 The antibody, or one of its functional fragments, according to the invention is also characterized in that it is able to inhibit the *in vivo* growth of tumor cell lines.

 The antibodies according to the present invention are preferably specific
30 monoclonal antibodies, especially of murine, chimeric or humanized origin, which can be obtained according to the standard methods well known to the person skilled in the art.

In general, for the preparation of monoclonal antibodies or their functional fragments, especially of murine origin, it is possible to refer to techniques which are described in particular in the manual "Antibodies" (Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor NY, pp. 726, 5 1988) or to the technique of preparation from hybridomas described by Kohler and Milstein (Nature, 256:495-497, 1975).

The monoclonal antibodies according to the invention can be obtained, for example, from an animal cell immunized against the IGF-IR, or one of its fragments containing the epitope specifically recognized by said monoclonal antibodies according 10 to the invention. Said IGF-IR, or one of its said fragments, can especially be produced according to the usual working methods, by genetic recombination starting with a nucleic acid sequence contained in the cDNA sequence coding for the IGF-IR or by peptide synthesis starting from a sequence of amino acids comprised in the peptide sequence of the IGF-IR.

15 The monoclonal antibodies according to the invention can, for example, be purified on an affinity column on which the IGF-IR or one of its fragments containing the epitope specifically recognized by said monoclonal antibodies according to the invention has previously been immobilized. More particularly, said monoclonal antibodies can be purified by chromatography on protein A and/or G, followed or not 20 followed by ion-exchange chromatography aimed at eliminating the residual protein contaminants as well as the DNA and the LPS, in itself followed or not followed by exclusion chromatography on Sepharose gel in order to eliminate the potential aggregates due to the presence of dimers or of other multimers. In an even more preferred manner, the whole of these techniques can be used simultaneously or 25 successively.

Chimeric or humanized antibodies are likewise included in antibodies according to the present invention.

By chimeric antibody, it is intended to indicate an antibody which contains a natural variable (light chain and heavy chain) region derived from an antibody of a 30 given species in combination with the light chain and heavy chain constant regions of an antibody of a species heterologous to said given species.

The antibodies or their fragments of chimeric type according to the invention can be prepared by using the techniques of genetic recombination. For example, the chimeric antibody can be produced by cloning a recombinant DNA containing a promoter and a sequence coding for the variable region of a nonhuman, especially murine, monoclonal antibody according to the invention and a sequence coding for the constant region of human antibody. A chimeric antibody of the invention encoded by such a recombinant gene will be, for example, a mouse-man chimera, the specificity of this antibody being determined by the variable region derived from the murine DNA and its isotype determined by the constant region derived from the human DNA. For the methods of preparation of chimeric antibodies, it is possible, for example, to refer to the document Verhoeyn et al. (BioEssays, 8:74, 1988).

By humanized antibody, it is intended to indicate an antibody which contains CDR regions derived from an antibody of nonhuman origin, the other parts of the antibody molecule being derived from one (or from several) human antibodies. Moreover, some of the residues of the segments of the skeleton (called FR) can be modified in order to conserve the affinity of the binding (Jones et al., Nature, 321:522-525, 1986; Verhoeyen et al., Science, 239:1534-1536, 1988; Riechmann et al., Nature, 332:323-327, 1988).

The humanized antibodies according to the invention or their fragments can be prepared by techniques known to the person skilled in the art (such as, for example, those described in the documents Singer et al., J. Immun., 150:2844-2857, 1992; Mountain et al., Biotechnol. Genet. Eng. Rev., 10:1-142, 1992; or Bebbington et al., Bio/Technology, 10:169-175, 1992). Such humanized antibodies according to the invention are preferred for their use in *in vitro* diagnostic methods, or *in vivo* prophylactic and/or therapeutic treatment. Other humanization methods are known by the man skill in the art as, for example, the "CDR Grafting" method described by Protein Design Lab (PDL) in the patent applications EP 0 451 261, EP 0 682 040, EP 0 9 127, EP 0 566 647 or US 5,530,101, US 6,180,370, US 5,585,089 and US, 5,693,761. The following patent applications can also be mentioned: US 5,639,641; US 6,054,297; US 5,886,152 and US 5,877,293. By functional fragment of an antibody according to the invention, it is intended to indicate in particular an antibody fragment, such as Fv, scFv (sc for single chain), Fab, F(ab')₂, Fab', scFv-Fc fragments or diabodies, or any

fragment of which the half-life time would have been increased by chemical modification, such as the addition of poly(alkylene) glycol such as poly(ethylene) glycol ("PEGylation") (pegylated fragments called Fv-PEG, scFv-PEG, Fab-PEG, F(ab')₂-PEG or Fab'-PEG) ("PEG" for Poly(Ethylene) Glycol), or by incorporation in a liposome, said fragments having at least one of the characteristic CDRs of sequence SEQ ID No. 1, 2, 3, 4, 5 or 6 according to the invention, and, especially, in that it is capable of exerting in a general manner an even partial activity of the antibody from which it is descended, such as in particular the capacity to recognize and to bind to the IGF-IR, and, if necessary, to inhibit the activity of the IGF-IR.

10 Preferably, said functional fragments will be constituted or will comprise a partial sequence of the heavy or light variable chain of the antibody from which they are derived, said partial sequence being sufficient to retain the same specificity of binding as the antibody from which it is descended and a sufficient affinity, preferably at least equal to 1/100, in a more preferred manner to at least 1/10, of that of the antibody from which it is descended, with respect to the IGF-IR.

Such a functional fragment will contain at the minimum 5 amino acids, preferably 10, 15, 25, 50 and 100 consecutive amino acids of the sequence of the antibody from which it is descended.

20 Preferably, these functional fragments will be fragments of Fv, scFv, Fab, F(ab')₂, F(ab'), scFv-Fc type or diabodies, which generally have the same specificity of binding as the antibody from which they are descended. According to the present invention, antibody fragments of the invention can be obtained starting from antibodies such as described above by methods such as digestion by enzymes, such as pepsin or papain and/or by cleavage of the disulfide bridges by chemical reduction. In another manner, the antibody fragments comprised in the present invention can be obtained by techniques of genetic recombination likewise well known to the person skilled in the art or else by peptide synthesis by means of, for example, automatic peptide synthesizers such as those supplied by the company Applied Biosystems, etc..

30 In a more preferred manner, the invention comprises the antibodies, or their functional fragments, according to the present invention, especially chimeric or humanized antibodies, obtained by genetic recombination or by chemical synthesis.

More particularly, according to a preferred embodiment of the invention, the antibody is characterized in that it comprises a light chain comprising at least one complementarity determining region CDR chosen from the CDRs of sequence SEQ ID No. 1, 3 or 5, or at least one CDR whose sequence has at least 80 % identity after optimum alignment with the sequence SEQ ID No. 1, 3 or 5, or in that it comprises a heavy chain comprising at least one CDR chosen from the CDRs of sequence SEQ ID No. 2, 4 or 6, or at least one CDR whose sequence has at least 80% identity after optimum alignment with the sequence SEQ ID No. 2, 4 or 6.

In the present description, the terms polypeptides, polypeptide sequences, peptides and proteins attached to antibody compounds or to their sequence are interchangeable.

It must be understood here that the invention does not relate to the antibodies in natural form, that is to say they are not in their natural environment but that they have been able to be isolated or obtained by purification from natural sources, or else obtained by genetic recombination, or by chemical synthesis, and that they can then contain unnatural amino acids as will be described further on.

By CDR region or CDR, it is intended to indicate the hypervariable regions of the heavy and light chains of the immunoglobulins as defined by Kabat et al. (Kabat et al., Sequences of proteins of immunological interest, 5th Ed., U.S. Department of Health and Human Services, NIH, 1991, and later editions). 3 heavy chain CDRs and 3 light chain CDRs exist. The term CDR or CDRs is used here in order to indicate, according to the case, one of these regions or several, or even the whole, of these regions which contain the majority of the amino acid residues responsible for the binding by affinity of the antibody for the antigen or the epitope which it recognizes.

By "percentage of identity" between two nucleic acid or amino acid sequences in the sense of the present invention, it is intended to indicate a percentage of nucleotides or of identical amino acid residues between the two sequences to be compared, obtained after the best alignment (optimum alignment), this percentage being purely statistical and the differences between the two sequences being distributed randomly and over their entire length. The comparisons of sequences between two nucleic acid or amino acid sequences are traditionally carried out by comparing these sequences after having aligned them in an optimum manner, said comparison being able

to be carried out by segment or by "comparison window". The optimum alignment of the sequences for the comparison can be carried out, in addition to manually, by means of the local homology algorithm of Smith and Waterman (1981) [Ad. App. Math. 2:482], by means of the local homology algorithm of Neddleman and Wunsch (1970) [J. Mol. Biol. 48: 443], by means of the similarity search method of Pearson and Lipman (1988) [Proc. Natl. Acad. Sci. USA 85:2444], by means of computer software using these algorithms (GAP, BESTFIT, FASTA and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI, or else by BLAST N or BLAST P comparison software).

10 The percentage of identity between two nucleic acid or amino acid sequences is determined by comparing these two sequences aligned in an optimum manner and in which the nucleic acid or amino acid sequence to be compared can comprise additions or deletions with respect to the reference sequence for an optimum alignment between these two sequences. The percentage of identity is calculated by determining the
15 number of identical positions for which the nucleotide or the amino acid residue is identical between the two sequences, by dividing this number of identical positions by the total number of positions in the comparison window and by multiplying the result obtained by 100 in order to obtain the percentage of identity between these two sequences.

20 For example, it is possible to use the BLAST program, "BLAST 2 sequences" (Tatusova et al., "Blast 2 sequences - a new tool for comparing protein and nucleotide sequences", FEMS Microbiol Lett. 174:247-250) available on the site <http://www.ncbi.nlm.nih.gov/gorf/bl2.html>, the parameters used being those given by default (in particular for the parameters "open gap penalty": 5, and "extension gap
25 penalty": 2; the matrix chosen being, for example, the matrix "BLOSUM 62" proposed by the program), the percentage of identity between the two sequences to be compared being calculated directly by the program.

By amino acid sequence having at least 80 %, preferably 85 %, 90 %, 95 % and 98 % identity with a reference amino acid sequence, those having, with respect to the
30 reference sequence, certain modifications, in particular a deletion, addition or substitution of at least one amino acid, a truncation or an elongation are preferred. In the case of a substitution of one or more consecutive or nonconsecutive amino acid(s), the

substitutions are preferred in which the substituted amino acids are replaced by “equivalent” amino acids. The expression “equivalent amino acids” is aimed here at indicating any amino acid capable of being substituted with one of the amino acids of the base structure without, however, essentially modifying the biological activities of the corresponding antibodies and such as will be defined later, especially in the examples.

Alternative embodiments of the invention propose anti-IGF-IR antibodies having other amino acid sequence modification(s) contained therein. For example, it may be desirable to improve the binding affinity and/or other biological properties of the antibody. Amino acid sequence variants of the anti-IGF-IR antibody are prepared by introducing appropriate nucleotide changes into the anti-IGF-IR antibody nucleic acid, or by peptide synthesis. Such modifications include, for example, deletions from, and/or insertions into and/or substitutions of, residues within the amino acid sequences of the anti-IGF-IR antibody. Any combination of deletion, insertion, and substitution is made to arrive at the final construct, provided that the final construct possesses the desired characteristics. The amino acid changes also may alter post-translational processes of the anti-IGF-IR antibody, such as changing the number or position of glycosylation sites.

A useful method for identification of certain residues or regions of the anti-IGF-IR antibody that are preferred locations for mutagenesis is called “alanine scanning mutagenesis” as described by Cunningham and Wells in Science, 244:1081-1085 (1989). Here, a residue or group of target residues are identified (e.g., charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids with IGF-IR antigen. Those amino acid locations demonstrating functional sensitivity to the substitutions then are refined by introducing further or other variants at, or for, the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation per se need not be predetermined. For example, to analyze the performance of a mutation at a given site, ala scanning or random mutagenesis is conducted at the target codon or region and the expressed anti-IGF-IR antibody variants are screened for the desired activity.

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Amino acid sequence insertions include amino- and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues. Examples of terminal insertions include an anti-IGF-IR antibody, with an N-terminal methionyl residue or the antibody fused to a cytotoxic polypeptide. Other insertional variants of the anti-IGF-IR antibody molecule include the fusion to the N- or C-terminus of the anti-IGF-IR antibody to an enzyme (e.g. for ADEPT) or a polypeptide which increases the serum half-life of the antibody.

Another type of variant is an amino acid substitution variant. These variants have at least one amino acid residue in the anti-IGF-IR antibody molecule replaced by a different residue. The sites of greatest interest for substitutional mutagenesis include the hypervariable regions, but FR alterations are also contemplated. Conservative substitutions are shown in Table 1 under the heading of "preferred substitutions". If such substitutions result in a change in biological activity, then more substantial changes, denominated "exemplary substitutions" in Table 1, or as further described below in reference to amino acid classes, may be introduced and the products screened.

Table 1**Amino Acid Substitutions**

<u>Original Residue</u>	<u>Exemplary Substitutions</u>	<u>Preferred Substitutions</u>
Ala (A)	val; leu; ile	val
Arg (R)	lys; gln; asn	lys
Asn (N)	gln; his; asp; lys; arg	gln
Asp (D)	glu; asn	glu
Cys (C)	ser; ala	ser
Gln (Q)	asn; glu	asn
Glu (E)	asp; gln	asp
Gly (G)	ala	ala
His (H)	asn; gln; lys; arg	arg
Ile (I)	leu; val; met; ala; phe; norleucine	leu
Leu (L)	norleucine; ile; val; met; ala; phe	ile
Lys (K)	arg; gln; asn	arg
Met (M)	leu; phe; ile	leu
Phe (F)	leu; val; ile; ala; tyr	tyr
Pro (P)	ala	ala
Ser (S)	thr	thr
Thr (T)	ser	ser
Trp (W)	tyr; phe	tyr
Tyr (Y)	trp; phe; thr; ser	phe
Val (V)	ile; leu; met; phe; ala; norleucine	leu

- 5 Substantial modifications in the biological properties of the antibody can be accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues
- 10 are divided into groups based on common side-chain properties:
- (1) hydrophobic: norleucine, met, ala, val, leu, ile;
 - (2) neutral hydrophilic: cys, ser, thr;
 - (3) acidic: asp, glu;
 - (4) basic: asn, gln, his, lys, arg;
 - 15 (5) residues that influence chain orientation: gly, pro; and
 - (6) aromatic: trp, tyr, phe.

Non-conservative substitutions will entail exchanging a member of one of these classes for another class.

Any cysteine residue not involved in maintaining the proper conformation of the anti-IGF-IR antibody also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking. Conversely, cysteine bond(s) may be added to the antibody to improve its stability (particularly where the antibody is an antibody fragment such as an Fv fragment).

A particularly preferred type of substitutional variant involves substituting one or more hypervariable region residues of a parent antibody (e.g. a humanized or human antibody). Generally, the resulting variant(s) selected for further development will have improved biological properties relative to the parent antibody from which they are generated. A convenient way for generating such substitutional variants involves affinity maturation using phage display. Briefly, several hypervariable region sites (e.g. 6-7 sites) are mutated to generate all possible amino substitutions at each site. The antibody variants thus generated are displayed in a monovalent fashion from filamentous phage particles as fusions to the gene III product of M13 packaged within each particle. The phage-displayed variants are then screened for their biological activity (e.g. binding affinity) as herein disclosed. In order to identify candidate hypervariable region sites for modification, alanine scanning mutagenesis can be performed to identify hypervariable region residues contributing significantly to antigen binding. Alternatively, or additionally, it may be beneficial to analyze a crystal structure of the antigen-antibody complex to identify contact points between the antibody and human IGF-IR. Such contact residues and neighboring residues are candidates for substitution according to the techniques elaborated herein. Once such variants are generated, the panel of variants is subjected to screening as described herein and antibodies with superior properties in one or more relevant assays may be selected for further development.

Another type of amino acid variant of the antibody alters the original glycosylation pattern of the antibody. By altering is meant deleting one or more carbohydrate moieties found in the antibody, and/or adding one or more glycosylation sites that are not present in the antibody.

Glycosylation of antibodies is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tripeptide sequences asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tripeptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-acetylgalactosamine, galactose, or xylose to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

Addition of glycosylation sites to the antibody is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tripeptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the sequence of the original antibody (for O-linked glycosylation sites).

Nucleic acid molecules encoding amino acid sequence variants of the subject anti-IGF-IR antibody are prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of the anti-IGF-IR antibody.

At times, it may be desirable to modify the antibody of the invention with respect to effector function, e.g. so as to enhance antigen-dependent cell-mediated cytotoxicity (ADCC) and/or complement dependent cytotoxicity (CDC) of the antibody. This may be achieved by introducing one or more amino acid substitutions in an Fc region of the antibody. Alternatively or additionally, cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron et al., *J. Exp Med.*, 176:1191-1195 (1992) and Shopes, B. *J. Immunol.*, 148:2918-2922 (1992). Homodimeric antibodies with enhanced anti-tumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff et al., *Cancer Research*, 53:2560-2565 (1993). Alternatively, an

antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson et al., Anti-Cancer Drug Design 3:219-230 (1989).

5 Likewise, to increase the serum half life of the antibody, one may incorporate a salvage receptor binding epitope into the antibody (especially an antibody fragment) as described in U.S. Pat. No. 5,739,277, for example. As used herein, the term "salvage receptor binding epitope" refers to an epitope of the Fc region of an IgG molecule (e.g., IgG₁, IgG₂, IgG₃, or IgG₄) that is responsible for increasing the *in vivo* serum half-life of the IgG molecule.

10 In a more preferred manner, the present invention relates to an antibody or one of its functional fragments, according to the invention, characterized in that it comprises a heavy chain comprising at least two of the three CDRs or the three CDRs of sequence SEQ ID Nos. 2, 4 and 6, or at least two of three CDRs or three CDRs of sequence respectively having at least 80 % identity after optimum alignment with the sequence
15 SEQ ID Nos. 2, 4 and 6.

In a more preferred embodiment, a subject of the invention is an antibody or one of its functional fragments according to the invention, characterized in that it comprises a light chain comprising at least two of the three CDRs or the three CDRs of sequence SEQ ID Nos. 1, 3 and 5, or at least two of three CDRs or three CDRs of sequence
20 respectively having at least 80% identity after optimum alignment with the sequence SEQ ID Nos. 1, 3 and 5.

In a more preferred manner, the antibody or one of its functional fragments according to the invention is characterized in that it comprises a heavy chain comprising the three CDRs of sequence SEQ ID Nos. 2, 4 and 6, or three CDRs of sequence
25 respectively having at least 80 % of identity after optimum alignment with the sequence SEQ ID Nos. 2, 4 and 6 and in that it moreover comprises a light chain comprising the three CDRs of sequence SEQ ID Nos. 1, 3 and 5, or three CDRs of sequence respectively having at least 80 % of identity after optimum alignment with the sequence SEQ ID Nos. 1, 3 and 5.

30 As state in the art, it is known that the largest variability (length and composition) between the 6 CDRs is observed for the 3 CDRs of the heavy chain and, more particularly, the CDR-H3. As a consequence, it will be understand that the

preferential characteristic CDR of the antibody of the invention is the 3 CDRs of the heavy chain, i.e. CDRs coded by sequence SEQ ID Nos. 2, 4 and 6 and, more preferentially, the CDR corresponding to CDR-H3 coded by SEQ ID No. 6.

According to another aspect, a subject of the present invention is an antibody or
5 one of its functional fragments, according to the invention, characterized in that it does not attach or it does not attach in a significant manner to the human insulin receptor IR.

In a preferred manner, said functional fragments according to the present invention will be chosen from the fragments Fv, scFv, Fab, (Fab')₂, Fab', scFv-Fc or diabodies, or any functional fragment whose half-life would have been increased by a
10 chemical modification, especially by PEGylation, or by incorporation in a liposome.

According to another aspect, the invention relates to a murine hybridoma capable of secreting a monoclonal antibody according to the present invention, especially the hybridoma of murine origin such as deposited at the Centre National de Culture De Microorganisme (CNCM, National Center of Microorganism Culture)
15 (Institut Pasteur, Paris, France) on June 23, 2005 under the number I-3466, said hybridoma resulting from the fusion of Balb/c splenocytes of immunized mice and Sp2O Ag 14 myeloma cell lines.

The monoclonal antibody here called I-3466, or one of its functional fragments, characterized in that said antibody is secreted by the hybridoma deposited at the CNCM
20 on June 23, 2005 under the number I-3466 is, of course, part of the present invention.

In a particular embodiment, the present invention relates to a murine antibody, or one of its functional fragments, according to the invention, characterized in that said antibody comprises a light chain of sequence comprising the amino acid sequence SEQ ID No. 7, or a sequence having at least 80% identity after optimum alignment with the
25 sequence SEQ ID No. 7, or/and in that it comprises a heavy chain of sequence comprising the amino acid sequence SEQ ID No. 8, or a sequence having at least 80 % identity after optimum alignment with the sequence SEQ ID No. 8.

According to a likewise particular aspect, the present invention relates to a chimeric antibody, or one of its functional fragments, according to the invention,
30 characterized in that said antibody moreover comprises the light chain and heavy chain constant regions derived from an antibody of a species heterologous to the mouse, especially man, and in a preferred manner in that the light chain and heavy chain

constant regions derived from a human antibody are respectively the kappa and gamma-1, gamma-2 or gamma-4 region.

In a particular embodiment corresponding to the isotype IgG1, a supplementary characteristic of the antibody is to potentially display effector functions as ADCC (for
5 Antibody Dependent Cellular Cytotoxicity) and/or CDC (for Complement Dependent Cytotoxicity).

According to a likewise particular aspect, the present invention relates to a humanized antibody or one of its functional fragments, according to the invention, characterized in that said antibody comprises a light chain and/or a heavy chain in
10 which the skeleton segments FR1 to FR4 of said light chain and/or heavy chain are respectively derived from skeleton segments FR1 to FR4 of human antibody light chain and/or heavy chain.

According to a preferred embodiment, the humanized antibody or one of its functional fragments, according to the present invention is characterized in that said
15 humanized antibody comprises a light chain comprising the amino acid sequence SEQ ID No. 17, or a sequence having at least 80% identity after optimum alignment with the sequence SEQ ID No. 17, or/and in that it comprises a heavy chain comprising the amino acid sequence SEQ ID No. 18, or a sequence having at least 80% identity after optimum alignment with the sequence SEQ ID No. 18.

20 Preferably, the humanized antibody, or one of its functional fragments, according to the invention is characterized in that said humanized antibody comprises a light chain comprising the amino acid sequence SEQ ID No. 17 and in that it comprises a heavy chain of sequence comprising the amino acid sequence SEQ ID No. 18.

According to a novel aspect, the present invention relates to an isolated nucleic
25 acid, characterized in that it is chosen from the following nucleic acids:

- a) a nucleic acid, DNA or RNA, coding for an antibody, or one of its functional fragments, according to the invention;
- b) a complementary nucleic acid of a nucleic acid such as defined in a);
- c) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions
30 of great stringency with at least one of the CDRs of nucleic acid sequence SEQ ID No. 9, 10, 11, 12, 13 or 14, or with a sequence having at least 80 %,

preferably 85 %, 90 %, 95 % and 98 %, identity after optimum alignment with the sequence SEQ ID No. 9, 10, 11, 12, 13 or 14;

d) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions of great stringency with at least the light chain of nucleic acid sequence SEQ ID No. 15 and/or the heavy chain of nucleic acid sequence SEQ ID No. 16, or with a sequence having at least 80 %, preferably 85 %, 90 %, 95 % and 98 %, identity after optimum alignment with the sequence SEQ ID No. 15 and/or 16;

e) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions of great stringency with at least the light chain of nucleic acid sequence SEQ ID No. 19 and/or the heavy chain of nucleic acid sequence SEQ ID No. 20, or with a sequence having at least 80 %, preferably 85 %, 90 %, 95 % and 98 %, identity after optimum alignment with the sequence SEQ ID No. 19 and/or 20.

By nucleic acid, nucleic or nucleic acid sequence, polynucleotide, oligonucleotide, polynucleotide sequence, nucleotide sequence, terms which will be employed indifferently in the present specification, it is intended to indicate a precise linkage of nucleotides, which are modified or unmodified, allowing a fragment or a region of a nucleic acid to be defined, containing or not containing unnatural nucleotides, and being able to correspond just as well to a double-stranded DNA, a single-stranded DNA as to the transcription products of said DNAs.

It must also be understood here that the present invention does not concern the nucleotide sequences in their natural chromosomal environment, that is to say in the natural state. It concerns sequences which have been isolated and/or purified, that is to say that they have been selected directly or indirectly, for example by copy, their environment having been at least partially modified. It is thus likewise intended to indicate here the isolated nucleic acids obtained by genetic recombination by means, for example, of host cells or obtained by chemical synthesis.

By nucleic sequences having a percentage of identity of at least 80 %, preferably 85 %, 90 %, 95 % and 98 %, after optimum alignment with a preferred sequence, it is intended to indicate the nucleic sequences having, with respect to the reference nucleic sequence, certain modifications such as, in particular, a deletion, a truncation, an elongation, a chimeric fusion and/or a substitution, especially point substitution. It preferably concerns sequences in which the sequences code for the same amino acid

sequences as the reference sequence, this being connected to the degeneracy of the genetic code, or complementary sequences which are capable of hybridizing specifically with the reference sequences, preferably under conditions of high stringency, especially such as defined below.

5 A hybridization under conditions of high stringency signifies that the temperature conditions and ionic strength conditions are chosen in such a way that they allow the maintenance of the hybridization between two fragments of complementary DNA. By way of illustration, conditions of high stringency of the hybridization step for the purposes of defining the polynucleotide fragments described above are
10 advantageously the following.

 The DNA-DNA or DNA-RNA hybridization is carried out in two steps: (1) prehybridization at 42°C for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5 x SSC (1 x SSC corresponds to a 0.15 M NaCl + 0.015 M sodium citrate solution), 50 % of formamide, 7 % of sodium dodecyl sulfate (SDS), 10 x Denhardt's, 5 % of
15 dextran sulfate and 1 % of salmon sperm DNA; (2) actual hybridization for 20 hours at a temperature dependent on the size of the probe (i.e.: 42°C, for a probe size > 100 nucleotides) followed by 2 washes of 20 minutes at 20°C in 2 x SSC + 2 % of SDS, 1 wash of 20 minutes at 20°C in 0.1 x SSC + 0.1 % of SDS. The last wash is carried out in 0.1 x SSC + 0.1 % of SDS for 30 minutes at 60°C for a probe size > 100 nucleotides.
20 The hybridization conditions of high stringency described above for a polynucleotide of defined size can be adapted by the person skilled in the art for oligonucleotides of greater or smaller size, according to the teaching of Sambrook et al., (1989, Molecular cloning: a laboratory manual. 2nd Ed. Cold Spring Harbor).

 The invention likewise relates to a vector comprising a nucleic acid according to
25 the present invention.

 The invention aims especially at cloning and/or expression vectors which contain a nucleotide sequence according to the invention.

 The vectors according to the invention preferably contain elements which allow the expression and/or the secretion of the nucleotide sequences in a determined host
30 cell. The vector must therefore contain a promoter, signals of initiation and termination of translation, as well as appropriate regions of regulation of transcription. It must be able to be maintained in a stable manner in the host cell and can optionally have

particular signals which specify the secretion of the translated protein. These different elements are chosen and optimized by the person skilled in the art as a function of the host cell used. To this effect, the nucleotide sequences according to the invention can be inserted into autonomous replication vectors in the chosen host, or be integrative vectors
5 of the chosen host.

Such vectors are prepared by methods currently used by the person skilled in the art, and the resulting clones can be introduced into an appropriate host by standard methods, such as lipofection, electroporation, thermal shock, or chemical methods.

The vectors according to the invention are, for example, vectors of plasmidic or
10 viral origin. They are useful for transforming host cells in order to clone or to express the nucleotide sequences according to the invention.

The invention likewise comprises the host cells transformed by or comprising a vector according to the invention.

The host cell can be chosen from prokaryotic or eukaryotic systems, for example
15 bacterial cells but likewise yeast cells or animal cells, in particular mammalian cells. It is likewise possible to use insect cells or plant cells.

The invention likewise concerns animals, except man, which comprise at least one cell transformed according to the invention.

According to another aspect, a subject of the invention is a process for
20 production of an antibody, or one of its functional fragments according to the invention, characterized in that it comprises the following stages:

- a) culture in a medium and appropriate culture conditions of a host cell according to the invention; and
- b) the recovery of said antibodies, or one of their functional fragments, thus
25 produced starting from the culture medium or said cultured cells.

The cells transformed according to the invention can be used in processes for preparation of recombinant polypeptides according to the invention. The processes for preparation of a polypeptide according to the invention in recombinant form, characterized in that they employ a vector and/or a cell transformed by a vector
30 according to the invention, are themselves comprised in the present invention. Preferably, a cell transformed by a vector according to the invention is cultured under

conditions which allow the expression of said polypeptide and said recombinant peptide is recovered.

As has been said, the host cell can be chosen from prokaryotic or eukaryotic systems. In particular, it is possible to identify nucleotide sequences according to the invention, facilitating secretion in such a prokaryotic or eukaryotic system. A vector according to the invention carrying such a sequence can therefore advantageously be used for the production of recombinant proteins, intended to be secreted. In effect, the purification of these recombinant proteins of interest will be facilitated by the fact that they are present in the supernatant of the cell culture rather than in the interior of the host cells.

It is likewise possible to prepare the polypeptides according to the invention by chemical synthesis. Such a preparation process is likewise a subject of the invention. The person skilled in the art knows the processes of chemical synthesis, for example the techniques employing solid phases (see especially Steward et al., 1984, Solid phase peptide synthesis, Pierce Chem. Company, Rockford, 111, 2nd ed., (1984)) or techniques using partial solid phases, by condensation of fragments or by a classical synthesis in solution. The polypeptides obtained by chemical synthesis and being able to contain corresponding unnatural amino acids are likewise comprised in the invention.

The antibodies, or one of their functional fragments, capable of being obtained by a process according to the invention are likewise comprised in the present invention.

According to a second embodiment, the present invention concerns an antibody according to the invention such as described further above, characterized in that it is, moreover, capable of binding specifically to a human receptor of the tyrosine kinase family and/or capable of inhibiting the tyrosine kinase activity of such receptor.

In a first embodiment, such an antibody consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the binding of EGF on the human epidermal growth factor receptor (EGFR) and/or inhibiting the tyrosine kinase activity of said EGFR. In a more preferred embodiment of the invention, said second anti-EGFR motif is issued from the murine monoclonal antibody 225, its chimeric derivative C225 or any humanized antibody derived from this antibody 225.

In a second embodiment, such an antibody consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the activity modulated by

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the HER2/neu receptor and/or inhibiting the tyrosine kinase activity of said HER2/neu receptor. In a more preferred embodiment of the invention, said second anti-HER2/neu motif is issued from the murine monoclonal antibody 4D5 or 2C4 or the humanized antibody Trastuzumab or Pertuzumab.

5 In a third embodiment, such an antibody consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the binding of the Hepatocyte Growth Factor (HGF) on the cMET receptor and/or specifically inhibiting the tyrosine kinase activity of said cMET receptor.

10 Finally, in a last embodiment, such an antibody consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the binding of the Macrophage Stimulating Protein (MSP) on the RON receptor and/or inhibiting the tyrosine kinase activity of said RON receptor.

15 According to another embodiment of the invention, it is also considered that the antibody according to the invention is capable to interact with any kind of receptor being implicated in the development of tumors such as, for example, without limitation, VEGFR, FGF (Fibroblast Growth Factor), PDGF (Platelet derived growth factor) or CXCR4 or 2 (Chemokine receptor type 4 or 2).

20 The bispecific or bifunctional antibodies form a second generation of monoclonal antibodies in which two different variable regions are combined in the same molecule (Hollinger and Bohlen, 1999, Cancer and metastasis, rev. 18:411-419). Their use has been demonstrated both in the diagnostic field and in the therapy field from their capacity to recruit new effector functions or to target several molecules on the surface of tumor cells. These antibodies can be obtained by chemical methods (Glennie MJ et al., 1987, J. Immunol., 139, 2367-2375; Repp R. et al., 1995, J. Hemat., 377-382)
25 or somatic methods (Staerz U.D. and Bevan M.J., 1986, PNAS 83, 1453-1457; Suresh M.R. et al., 1986, Method Enzymol., 121:210-228) but likewise and preferentially by genetic engineering techniques which allow the heterodimerization to be forced and thus facilitate the process of purification of the antibody sought (Merchand et al., 1998 Nature Biotech., 16:677-681).

30 These bispecific antibodies can be constructed as entire IgG, as bispecific Fab'2, as Fab'PEG or as diabodies or else as bispecific scFv but likewise as a tetravalent

bispecific antibody or two attachment sites are present for each antigen targeted (Park et al., 2000, Mol. Immunol., 37 (18):1123-30) or its fragments as described further above.

In addition to an economic advantage from the fact that the production and the administration of a bispecific antibody are less onerous than the production of two specific antibodies, the use of such bispecific antibodies has the advantage of reducing the toxicity of the treatment. This is because the use of a bispecific antibody allows the total quantity of circulating antibodies to be reduced and, consequently, the possible toxicity.

In a preferred embodiment of the invention, the bispecific antibody is a bivalent or tetravalent antibody.

The invention likewise concerns a pharmaceutical composition comprising by way of active principle a compound consisting of an antibody, or one of its functional fragments according to the invention, preferably mixed with an excipient and/or a pharmaceutically acceptable vehicle.

According to yet another embodiment, the present invention likewise concerns a pharmaceutical composition such as described further above which comprises at least a second compound chosen from the compounds capable of specifically inhibiting the tyrosine kinase activity of IGF-IR, EGFR, HER2/neu, VEGFR, cMET and/or RON.

In a second preferred aspect of the invention, said second compound is chosen from the isolated anti-EGFR, -IGF-IR, -HER2/neu, anti-VEGFR, -cMET and/or -RON antibodies, or their functional fragments, capable of inhibiting the proliferative and/or anti-apoptotic and/or angiogenic and/or metastatic dissemination inductor activity mediated by said receptor(s).

According to another embodiment of the invention, the composition comprises, as combination product for a simultaneous, separate or sequential use, at least an inhibitor of the tyrosine kinase activity of the IGF-IR, EGFR, HER2/neu, VEGFR, cMET and/or RON.

In another preferred embodiment, said inhibitor of the tyrosine kinase activity of the receptors is selected from the group consisting of derived natural agents, dianilinophthalimides, pyrazolo- or pyrrolopyridopyrimidines or else quinazilines. Such inhibitory agents are well known to the person skilled in the art and described in the literature (Ciardiello F., Drugs 2000, Suppl. 1, 25-32).

Another complementary embodiment of the invention consists in a composition such as described above which comprises, moreover, as a combination product for simultaneous, separate or sequential use, a cytotoxic/cytostatic agent.

5 “Simultaneous use” is understood as meaning the administration of the two compounds of the composition according to the invention in a single and identical pharmaceutical form.

“Separate use” is understood as meaning the administration, at the same time, of the two compounds of the composition according to the invention in distinct pharmaceutical forms.

10 “Sequential use” is understood as meaning the successive administration of the two compounds of the composition according to the invention, each in a distinct pharmaceutical form.

In a general fashion, the composition according to the invention considerably increases the efficacy of the treatment of cancer. In other words, the therapeutic effect
15 of the anti-IGF-IR antibody according to the invention is potentiated in an unexpected manner by the administration of a cytotoxic agent. Another major subsequent advantage produced by a composition according to the invention concerns the possibility of using lower efficacious doses of active principle, which allows the risks of appearance of secondary effects to be avoided or to be reduced, in particular the effects of the
20 cytotoxic agent.

In addition, this composition according to the invention would allow the expected therapeutic effect to be attained more rapidly.

By “anti-cancer therapeutic agents” or “cytotoxic agents”, it is intended a substance which, when administered to a subject, treats or prevents the development of
25 cancer in the subject's body. As non limitative example of such agents, it can be mentioned alkylating agents, anti-metabolites, anti-tumor antibiotics, mitotic inhibitors, chromatin function inhibitors, anti-angiogenesis agents, anti-estrogens, anti-androgens or immunomodulators.

Such agents are, for example, cited in the 2001 edition of VIDAL, on the page
30 devoted to the compounds attached to the cancerology and hematology column “Cytotoxics”, these cytotoxic compounds cited with reference to this document are cited here as preferred cytotoxic agents.

More particularly, the following agents are preferred according to the invention.

“Alkylating agent” refers to any substance which can cross-link or alkylate any molecule, preferably nucleic acid (e.g., DNA), within a cell. Examples of alkylating agents include nitrogen mustard such as mechlorethamine, chlorambucol, melphalen, 5 chlorydrate, pipobromen, prednimustin, disodic-phosphate or estramustine; oxazophorins such as cyclophosphamide, altretamine, trofosfamide, sulfofosfamide or ifosfamide; aziridines or imine-ethylènes such as thiotepa, triethylenamine or altetramine; nitrosourea such as carmustine, streptozocin, fotemustin or lomustine; alkyle-sulfonates such as busulfan, treosulfan or improsulfan; triazenes such as 10 dacarbazine; or platinum complexes such as cis-platinum, oxaliplatin and carboplatin.

“Anti-metabolites” refer to substances that block cell growth and/or metabolism by interfering with certain activities, usually DNA synthesis. Examples of anti-metabolites include methotrexate, 5-fluoruracil, floxuridine, 5-fluorodeoxyuridine, capecitabine, cytarabine, fludarabine, cytosine arabinoside, 6-mercaptopurine (6-MP), 15 6-thioguanine (6-TG), chlorodesoxyadenosine, 5-azacytidine, gemcitabine, cladribine, deoxycoformycin and pentostatin.

“Anti-tumor antibiotics” refer to compounds which may prevent or inhibit DNA, RNA and/or protein synthesis. Examples of anti-tumor antibiotics include doxorubicin, daunorubicin, idarubicin, valrubicin, mitoxantrone, dactinomycin, mithramycin, 20 plicamycin, mitomycin C, bleomycin, and procarbazine.

“Mitotic inhibitors” prevent normal progression of the cell cycle and mitosis. In general, microtubule inhibitors or taxoides such as paclitaxel and docetaxel are capable of inhibiting mitosis. Vinca alkaloid such as vinblastine, vincristine, vindesine and vinorelbine are also capable of inhibiting mitosis.

25 “Chromatin function inhibitors” or “topoisomerase inhibitors” refer to substances which inhibit the normal function of chromatin modeling proteins such as topoisomerase I or topoisomerase II. Examples of chromatin function inhibitors include, for topoisomerase I, camptothecine and its derivatives such as topotecan or irinotecan, and, for topoisomerase II, etoposide, etoposide phosphate and teniposide.

30 “Anti-angiogenesis agent” refers to any drug, compound, substance or agent which inhibits growth of blood vessels. Exemplary anti-angiogenesis agents include, but are by no means limited to, razoxin, marimastat, batimastat, prinomastat, tanomastat,

ilomastat, CGS-27023A, halofuginon, COL-3, neovastat, BMS-275291, thalidomide, CDC 501, DMXAA, L-651582, squalamine, endostatin, SU5416, SU6668, interferon-alpha, EMD121974, interleukin-12, IM862, angiostatin and vitaxin.

“Anti-estrogen” or “anti-estrogenic agent” refer to any substance which reduces, antagonizes or inhibits the action of estrogen. Examples of anti-estrogen agents are tamoxifen, toremifene, raloxifene, droloxifene, iodoxyfene, anastrozole, letrozole, and exemestane.

“Anti-androgens” or “anti-androgen agents” refer to any substance which reduces, antagonizes or inhibits the action of an androgen. Examples of anti-androgens are flutamide, nilutamide, bicalutamide, sprironolactone, cyproterone acetate, finasteride and cimitidine.

“Immunomodulators” are substances which stimulate the immune system.

Examples of immunomodulators include interferon, interleukin such as aldesleukine, OCT-43, denileukin diflitox and interleukin-2, tumoral necrosis factors such as tasonermin or others immunomodulators such as lentinan, sizofiran, roquinimex, pidotimod, pegademase, thymopentine, poly I:C or levamisole in conjunction with 5-fluorouracil.

For more detail, the man skilled in the art could refer to the manual edited by the “Association Française des Enseignants de Chimie Thérapeutique” and entitled “traité de chimie thérapeutique, vol. 6, Médicaments antitumoraux et perspectives dans le traitement des cancers, edition TEC & DOC, 2003”.

In a particularly preferred embodiment, said composition as a combination product according to the invention is characterized in that said cytotoxic agent is coupled chemically to said antibody for simultaneous use.

In a particularly preferred embodiment, said composition according to the invention is characterized in that said cytotoxic/cytostatic agent is chosen from the spindle inhibitor or stabilizer agents, preferably vinorelbine and/or vinflunine and/or vincristine.

In order to facilitate the coupling between said cytotoxic agent and said antibody according to the invention, it is especially possible to introduce spacer molecules between the two compounds to be coupled, such as poly(alkylene) glycols like polyethylene glycol, or else amino acids, or, in another embodiment, to use active

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derivatives of said cytotoxic agents into which would have been introduced functions capable of reacting with said antibody according to the invention. These coupling techniques are well known to the person skilled in the art and will not be expanded upon in the present description.

5 Other inhibitors of EGFR can, without any limitation, consist of the anti-EGFR monoclonal antibodies C225 and 22Mab (ImClone Systems Incorporated), ABX-EGF (Abgenix/Cell Genesys), EMD-7200 (Merck KgaA) or the compounds ZD-1834, ZD-1838 and ZD-1839 (AstraZeneca), PKI-166 (Novartis), PKI-166/CGP-75166 (Novartis), PTK 787 (Novartis), CP 701 (Cephalon), leflunomide (Pharmacia/Sugen),
10 CI-1033 (Warner-Lambert Parke-Davis), CI-1033/PD 183, 805 (Warner-Lambert Parke-Davis), CL-387, 785 (Wyeth-Ayerst), BBR-1611 (Boehringer Mannheim GmbH/Roche), Naamidine A (Bristol-Myers Squibb), RC-3940-II (Pharmacia), BIBX-1382 (Boehringer Ingelheim), OLX-103 (Merck & Co), VRCTC-310 (Ventech Research), EGF fusion toxin (Seragen Inc.), DAB-389 (Seragen/Lilgand), ZM-252808
15 (Imperial Cancer Research Fund), RG-50864 (INSERM), LFM-A12 (Parker Hughes Cancer Center), WHI-P97 (Parker Hughes Cancer Center), GW-282974 (Glaxo), KT-8391 (Kyowa Hakko) or the "EGFR Vaccine" (York Medical/Centro de Immunologia Molecular).

 According to yet another embodiment of the invention, the composition such as
20 described above can likewise comprise another antibody compound directed against the extracellular domain of the HER2/neu receptor, as a combination product for simultaneous, separate or sequential use, intended for the prevention and for the treatment of cancer, especially the cancers overexpressing said HER2/neu receptor and the receptor IGF-IR and/or EGFR, such as especially cancer of the breast.

25 Reference can be made especially to the publications of Albanell et al. (J. of the National Cancer Institute, 93(24):1830-1831, 2001) and of Lu et al. (J. of the National Cancer Institute, 93(24):1852-1857, 2001) justifying the unexpected interest in combining an anti-HER2/neu antibody with an anti-IGF-IR antibody according to the present invention.

30 In a particular manner, said anti-HER2/neu antibody of the composition according to the invention is the antibody called Trastuzumab (also called Herceptin).

The invention relates, in another aspect, to a composition characterized in that one, at least, of said antibodies, or one of their functional fragments, is conjugated with a cell toxin and/or a radioelement.

Preferably, said toxin or said radioelement is capable of inhibiting at least one
5 cell activity of cells expressing the IGF-IR, in a more preferred manner capable of preventing the growth or the proliferation of said cell, especially of totally inactivating said cell.

Preferably also, said toxin is an enterobacterial toxin, especially *Pseudomonas* exotoxin A.

10 The radioelements (or radioisotopes) preferably conjugated to the antibodies employed for the therapy are radioisotopes which emit gamma rays and preferably iodine¹³¹, yttrium⁹⁰, gold¹⁹⁹, palladium¹⁰⁰, copper⁶⁷, bismuth²¹⁷ and antimony²¹¹. The radioisotopes which emit beta and alpha rays can likewise be used for the therapy.

By toxin or radioelement conjugated to at least one antibody, or one of its
15 functional fragments, according to the invention, it is intended to indicate any means allowing said toxin or said radioelement to bind to said at least one antibody, especially by covalent coupling between the two compounds, with or without introduction of a linking molecule.

Among the agents allowing binding in a chemical (covalent), electrostatic or
20 noncovalent manner of all or part of the components of the conjugate, mention may particularly be made of benzoquinone, carbodiimide and more particularly EDC (1-ethyl-3-[3-dimethylaminopropyl]-carbodiimide hydrochloride), dimaleimide, dithiobis-nitrobenzoic acid (DTNB), N-succinimidyl S-acetyl thio-acetate (SATA), the bridging agents having one or more phenylazide groups reacting with the ultraviolets (U.V.) and
25 preferably N-[-4-(azidosalicylamino)butyl]-3'-(2'-pyridyldithio)propionamide (APDP), N-succinimid-yl 3-(2-pyridyldithio)propionate (SPDP), 6-hydrazino-nicotinamide (HYNIC).

Another form of coupling, especially for the radioelements, can consist in the use of a bifunctional ion chelator.

30 Among these chelates, it is possible to mention the chelates derived from EDTA (ethylenediaminetetraacetic acid) or from DTPA (diethylenetriaminepentaacetic acid) which have been developed for binding metals, especially radioactive metals, and

immunoglobulins. Thus, DTPA and its derivatives can be substituted by different groups on the carbon chain in order to increase the stability and the rigidity of the ligand-metal complex (Krejcarek et al., 1977; Brechbiel et al., 1991; Gansow, 1991; US patent 4,831,175).

5 For example diethylenetriaminepentaacetic acid (DTPA) and its derivatives, which have been widely used in medicine and in biology for a long time either in their free form, or in the form of a complex with a metallic ion, have the remarkable characteristic of forming stable chelates with metallic ions and of being coupled with proteins of therapeutic or diagnostic interest such as antibodies for the development of
10 radioimmunoconjugates in cancer therapy (Meases et al., 1984; Gansow et al., 1990).

Likewise preferably, said at least one antibody forming said conjugate according to the invention is chosen from its functional fragments, especially the fragments amputated of their Fc component such as the scFv fragments.

15 The present invention moreover comprises the use of the composition according to the invention for the preparation of a medicament.

More particularly, according to another embodiment, the invention concerns the use of an antibody, or one of its functional fragments, and/or of a composition for the preparation of a medicament intended for the prevention or for the treatment of an illness induced by an overexpression and/or an abnormal activation of the IGF-IR, and/or connected with a hyperactivation of the transduction pathway of the signal
20 mediated by the interaction of the IGF1 or IGF2 with IGF-IR.

According to yet another preferred embodiment, the invention concerns the use of an antibody, or one of its functional fragments, and/or of a composition for the preparation of a medicament intended for the prevention or for the treatment of an illness induced by an overexpression and/or an abnormal activation of the IGF-IR, and/or connected with a hyperactivation of the transduction pathway of the signal
25 mediated by the interaction of the IGF2 with IGF-IR.

Preferably, said use according to the invention is characterized in that the administration of said medicament does not induce or induces only slightly secondary
30 effects connected with inhibition of the insulin receptor IR, that is to say inhibition of the interaction of the IR with its natural ligands due to the presence of said medicament,

especially by a competitive inhibition connected with the attachment of said medicament to the IR.

The present invention moreover comprises the use of an antibody, or one of its functional fragments, preferably humanized, and/or of a composition according to the invention for the preparation of a medicament intended to inhibit the transformation of
5 normal cells into cells with tumoral character, preferably IGF-dependent, especially at least IGF2-dependent cells.

The present invention likewise relates to the use of an antibody, or one of its functional fragments, preferably humanized, and/or of a composition according to the
10 invention for the preparation of a medicament intended to inhibit the growth and/or the proliferation of tumor cells, preferably IGF-dependent, especially at least IGF2-dependent cells.

In a general manner, a subject of the present invention is the use of an antibody, or one of its functional fragments, preferably humanized, and/or of a composition
15 according to the invention, for the preparation of a medicament intended for the prevention or for the treatment of cancer preferably expressing IGF-IR, and/or of cancer preferably having a hyperactivation of the transduction pathway of the signal mediated by the interaction of IGF1 and/or IGF2 with IGF-IR, such as, for example, the overexpression of IRS1.

The subject of the present invention is likewise the use of an antibody, or one of its functional fragments, preferably humanized, and/or of a composition according to the invention, for the preparation of a medicament intended for the prevention or for the treatment of psoriasis, psoriasis whose epidermal hyperproliferation can be connected with the expression or the overexpression of IGF-IR, and/or with the hyperactivation of
20 the transduction pathway of the signal mediated by the interaction of IGF-IR with its natural ligands (Wraight C.J. et al., Nat. Biotechnol., 2000, 18(5):521-526, Reversal of epidermal hyperproliferation in psoriasis by insulin-like growth factor I receptor antisense oligonucleotides) and/or of EGFR with its natural ligands.

The invention also relates to the use of the antibody, or any functional fragments thereof, preferentially humanized, and/or of any composition comprising said antibody,
30 for the preparation of a medicament for the treatment or the prevention of atherosclerosis.

Among the cancers which can be prevented and/or treated, prostate cancer, osteosarcomas, lung cancer, breast cancer, endometrial cancer, colon cancer, multiple myeloma or ovary cancer or any other cancer overexpressing IGF-IR is preferred.

5 In a more preferred embodiment, due to the particular ability to displace IGF2, the invention relates to the use of the antibody, or fragment thereof, according to the invention, for the prevention, diagnostic or treatment of colon cancer, as IGF2 is known to be particularly implicated in this cancer.

According to yet another aspect, a subject of the present invention is a method of diagnosis, preferably *in vitro*, of illnesses connected with an overexpression or an
10 underexpression, preferably an overexpression, of the IGF-IR starting from a biological sample in which the abnormal presence of IGF-IR is suspected, characterized in that said biological sample is contacted with an antibody, or one of its functional fragments, according to the invention, it being possible for said antibody to be, if necessary, labeled.

15 Preferably, said illnesses connected with the overexpression of the IGF-IR in said diagnosis method will be cancers.

Said antibody, or one of its functional fragments, can be present in the form of an immunoconjugate or of a labeled antibody so as to obtain a detectable and/or quantifiable signal.

20 An object of the invention is a method of *in vitro* diagnosis of pathological condition characterized by aberrant expression of IGF-IR relative to normal, said method comprising contacting a biological sample suspected of containing IGF-IR, with the antibody according the invention, under conditions favoring formation of an IGF-IR/antibody complex, and detecting said complex as indicating presence said IGF-IR in
25 said sample. In a preferred embodiment, said antibody is detectably labeled.

In a first aspect, the aberrant expression of IGF-IR is an over-expression of IGF-IR. In a second aspect, the aberrant expression of IGF-IR is an under-expression of IGF-IR.

The antibodies labeled according to the invention or their functional fragments
30 include, for example, antibodies called immunoconjugates which can be conjugated, for example, with enzymes such as peroxidase, alkaline phosphatase, α -D-galactosidase, glucose oxydase, glucose amylase, carbonic anhydrase, acetylcholinesterase, lysozyme,

malate dehydrogenase or glucose 6-phosphate dehydrogenase or by a molecule such as biotin, digoxigenin or 5-bromodeoxyuridine. Fluorescent labels can be likewise conjugated to the antibodies or to their functional fragments according to the invention and especially include fluorescein and its derivatives, fluorochrome, rhodamine and its derivatives, GFP (GFP for "Green Fluorescent Protein"), dansyl, umbelliferone etc.. In such conjugates, the antibodies of the invention or their functional fragments can be prepared by methods known to the person skilled in the art. They can be coupled to the enzymes or to the fluorescent labels directly or by the intermediary of a spacer group or of a linking group such as a polyaldehyde, like glutaraldehyde, ethylenediaminetetraacetic acid (EDTA), diethylene-triaminepentaacetic acid (DTPA), or in the presence of coupling agents such as those mentioned above for the therapeutic conjugates. The conjugates containing labels of fluorescein type can be prepared by reaction with an isothiocyanate.

Other conjugates can likewise include chemoluminescent labels such as luminol and the dioxetanes, bio-luminescent labels such as luciferase and luciferin, or else radioactive labels such as iodine¹²³, iodine¹²⁵, iodine¹²⁶, iodine¹³³, bromine⁷⁷, technetium^{99m}, indium¹¹¹, indium^{113m}, gallium⁶⁷, gallium⁶⁸, ruthenium⁹⁵, ruthenium⁹⁷, ruthenium¹⁰³, ruthenium¹⁰⁵, mercury¹⁰⁷, mercury²⁰³, rhenium^{99m}, rhenium¹⁰¹, rhenium¹⁰⁵, scandium⁴⁷, tellurium^{121m}, tellurium^{122m}, tellurium^{125m}, thulium¹⁶⁵, thulium¹⁶⁷, thulium¹⁶⁸, fluorine¹⁸, yttrium¹⁹⁹, iodine¹³¹. The methods known to the person skilled in the art existing for coupling the therapeutic radioisotopes to the antibodies either directly or via a chelating agent such as EDTA, DTPA mentioned above can be used for the radioelements which can be used in diagnosis. It is likewise possible to mention labeling with Na[I¹²⁵] by the chloramine T method [Hunter W.M. and Greenwood F.C. (1962) Nature 194:495] or else with technetium^{99m} by the technique of Crockford et al. (US patent 4,424,200) or attached via DTPA as described by Hnatowich (US patent 4,479,930).

Thus, the antibodies, or their functional fragments, according to the invention can be employed in a process for the detection and/or the quantification of an overexpression or of an underexpression, preferably an overexpression, of the IGF-IR in a biological sample, characterized in that it comprises the following steps:

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a) the contacting of the biological sample with an antibody, or one of its functional fragments, according to the invention; and

b) the demonstration of the IGF-IR /antibody complex possibly formed.

In a particular embodiment, the antibodies, or their functional fragments, according to the invention, can be employed in a process for the detection and/or the quantification of the IGF-IR in a biological sample, for the monitoring of the efficacy of a prophylactic and/or therapeutic treatment of IGF-dependent cancer or else of psoriasis or atherosclerosis.

More generally, the antibodies, or their functional fragments, according to the invention can be advantageously employed in any situation where the expression of the IGF-IR must be observed in a qualitative and/or quantitative manner.

Preferably, the biological sample is formed by a biological fluid, such as serum, whole blood, cells, a tissue sample or biopsies of human origin.

Another aspect of the invention is a diagnostic method for predicting an oncogenic potential of a sample of prostate cells, comprising:

(a) providing a sample of human prostate tissue; and

(b) determining, in the sample, presence of IGF-IR, said step comprising contacting said sample with the antibody according to the invention under conditions favoring formation of an IGF-IR/antibody complex, wherein presence of said complex indicates the oncogenic potential of said cells in said tissue.

In another embodiment, the presence of said complex indicates that the patient is at risk of developing or onset of a pathological disorder characterized by overexpression of said IGF-IR.

An object of the invention is also a method for following progress of a therapeutic regime designed to alleviate a pathological disorder characterized by abnormal expression of IGF-IR expression, comprising:

(a) assaying a sample from a subject to determine level of IGF-IR at a first time point;

(b) assaying said sample at a second time point; and

(c) comparing said level at said second time point to the level determined in (a) as a determination of effect of said therapeutic régime wherein a decrease in the level of IGF-IR in said sample is determinative of the regression said pathological disorder in

said patient or an increase in the level of IGF-IR is determinative of the progression of said pathological disorder in said patient.

Any procedure or conventional test can be employed in order to carry out such a detection and/or dosage. Said test can be a competition or sandwich test, or any test
5 known to the person skilled in the art dependent on the formation of an immune complex of antibody-antigen type. Following the applications according to the invention, the antibody or one of its functional fragments can be immobilized or labeled. This immobilization can be carried out on numerous supports known to the person skilled in the art. These supports can especially include glass, polystyrene, poly-
10 propylene, polyethylene, dextran, nylon, or natural or modified cells. These supports can be either soluble or insoluble.

By way of example, a preferred method brings into play immunoenzymatic processes according to the ELISA technique, by immunofluorescence, or radio-immunoassay (RIA) technique or equivalent.

15 Thus, the present invention likewise comprises the kits or sets necessary for carrying out a method of diagnosis of illnesses induced by an overexpression or an underexpression of the IGF-IR or for carrying out a process for the detection and/or the quantification of an overexpression or of an underexpression of the IGF-IR in a biological sample, preferably an overexpression of said receptor, characterized in that
20 said kit or set comprises the following elements:

- a) an antibody, or one of its functional fragments, according to the invention;
- b) optionally, the reagents for the formation of the medium favorable to the immunological reaction;
- c) optionally, the reagents allowing the demonstration of IGF-IR/antibody
25 complexes produced by the immunological reaction.

The invention moreover relates to the use of a composition as a combination product according to the invention, for the preparation of a medicament intended for the prevention or for the treatment of cancer, especially cancers for which said cytotoxic agent or said anti-HER2/neu antibody is generally prescribed and, especially, for which
30 cancers the tumor cells express or overexpress the IGF-IR.

A subject of the invention is likewise the use of an antibody according to the invention for the preparation of a medicament intended for the specific targeting of a biologically active compound to cells expressing or overexpressing the IGF-IR.

5 It is intended here by biologically active compound to indicate any compound capable of modulating, especially of inhibiting, cell activity, in particular their growth, their proliferation, transcription or gene translation.

A subject of the invention is also an *in vivo* diagnostic reagent comprising an antibody according to the invention, or one of its functional fragments, preferably labeled, especially radiolabeled, and its use in medical imaging, in particular for the
10 detection of cancer connected with the expression or the overexpression by a cell of the IGF-IR.

The invention likewise relates to a composition as a combination product or to an anti-IGF-IR/toxin conjugate or radioelement, according to the invention, as a medicament.

15 Preferably, said composition as a combination product or said conjugate according to the invention will be mixed with an excipient and/or a pharmaceutically acceptable vehicle.

In the present description, pharmaceutically acceptable vehicle is intended to indicate a compound or a combination of compounds entering into a pharmaceutical
20 composition not provoking secondary reactions and which allows, for example, facilitation of the administration of the active compound(s), an increase in its lifespan and/or in its efficacy in the body, an increase in its solubility in solution or else an improvement in its conservation. These pharmaceutically acceptable vehicles are well known and will be adapted by the person skilled in the art as a function of the nature
25 and of the mode of administration of the active compound(s) chosen.

Preferably, these compounds will be administered by the systemic route, in particular by the intravenous route, by the intramuscular, intradermal, intraperitoneal or subcutaneous route, or by the oral route. In a more preferred manner, the composition comprising the antibodies according to the invention will be administered several times,
30 in a sequential manner.

Their modes of administration, dosages and optimum pharmaceutical forms can be determined according to the criteria generally taken into account in the establishment

of a treatment adapted to a patient such as, for example, the age or the body weight of the patient, the seriousness of his/her general condition, the tolerance to the treatment and the secondary effects noted.

Other characteristics and advantages of the invention appear in the continuation
5 of the description with the examples and the figures whose legends are represented below.

LEGENDS TO THE FIGURES

Figure 1: Competition of [125 I]-IGF-1 binding to IGF-IR by the monoclonal antibody I-
10 3466.

Specific [125 I]-IGF-1 binding (in %) was plotted as a function of ligand concentration on a semilog graph. Specific binding values are the means of experiments performed in triplicate.

Figure 2: Competition of [125 I]-IGF-2 binding to IGF-IR by the monoclonal antibody I-
15 3466.

Specific [125 I]-IGF-2 binding (in %) was plotted as a function of ligand concentration on a semilog graph. Specific binding values are the means of experiments performed in triplicate.

Figures 3A and 3B: *In vitro* effect of the I-3466 antibody on either IGF1- or IGF2-
20 induced MCF-7 growth.

Figures 4A and 4B: Effect of the I-3466 Mab on either IGF1 (Figure 4A) or IGF2 (Figure 4B)-induced phosphorylation of IGF-IR β -chain on MCF-7 cells.

Figures 5A and 5B: Effect of the I-3466 Mab on either IGF1 (Figure 5A) or IGF2 (Figure 5B)-induced phosphorylation of IGF-IR β -chain on HT29 cells.

25 Figures 6A-6C: *In vivo* activity of I-3466 in DU145 (Figure 6A), SK-ES-1 (Figure 6B), HT29 (Figure 6C), A549 (Figure 6D) and MCF-7 (Figure 6E) xenograft tumour models.

Figure 7: Comparison of the amino acid sequences of the variable regions of the light chains (VL) of mouse I-3466 (SEQ ID No. 7) and of humanized by CDR-grafting I-3466 (SEQ ID No. 20).

30 The symbol * (asterisk) indicates important residues for the maintenance of CDR loop conformation and the symbol # (sharp) indicates conserved residues found at the VL/VH interface.

Figure 8: Comparison of the amino acid sequences of the variable regions of the heavy chains (VH) of mouse I-3466 (SEQ ID No. 8) and of humanized by CDR-grafting I-3466 (SEQ ID No. 21).

The symbol * (asterisk) indicates important residues for the maintenance of CDR loop conformation and the symbol # (sharp) indicates conserved residues found at the VL/VH interface.

Figure 9: SDS-PAGE analysis of purified I-3466 antibodies. The legend is the following:

M: Marker,
 10 Lane 1: I-3466 humanized variable region,
 Lane 2: I-3466 mouse variable region,
 Panel A: Reducing SDS-PAGE,
 Panel B: Non-reducing SDS-PAGE.

The constant regions for both mouse and humanized 3466 are human.

15 Figure 10: Schematic representation of the biosensor capturing assay.

A human IgG1 Mab directed against the constant Fc portion, was covalently attached onto a CM5 sensor surface. A limited amount of Mab to be tested was immobilized and used to capture the analyte hIGF-IR-ECD. The binding of Mab to the analyte is characterized by the association and dissociation rate constants k_a and k_d ,
 20 respectively. The equilibrium dissociation constant (KD) is calculated by the ratio between dissociation and association rate constants.

Figure 11: Sensorgram of the association and dissociation phase of IgG1 humanized I-3466 / hIGF-IR-ECD complexes for five different concentrations of hIGF-IR-ECD.

25 **Example 1: Generation and selection of the murine monoclonal antibody (MAb)**

With the aim of generating MAb specifically directed against IGF-IR and not recognizing the IR, a protocol comprising 6 screening stages was performed.

It consisted in:

- immunizing mice with human recombinant IGF-IR, in order to generate
 30 hybridomas,
- screening the culture supernatants by ELISA on the recombinant protein which served for immunization,
- testing all the supernatants of hybridomas positive by ELISA on the native receptor overexpressed on the surface of MCF-7 tumor cells,

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- performing binding experiments to select antibodies recognizing specifically IGF-IR,

- verifying that the antibodies selected at this stage were capable of inhibiting *in vitro* the induced IGF1 proliferation of the MCF-7 cells,

5 - ensuring the *in vivo* activity, in nude mice, of the candidate retained in terms of impact on the growth of the tumor MCF-7.

All of these different stages and results obtained will be briefly described below in example 1.

10 For the immunization stage, mice were injected twice, by the subcutaneous route, with 8 µg of recombinant IGF-IR. Three days before the fusion of spleen cells with the cells of the murine myeloma Sp2OAg14, the mice received by an intravenous injection of 3 µg of the recombinant receptor. Fourteen days after the fusion, the supernatants of hybridomas were screened by ELISA, on plates sensitized by recombinant IGF-IR. The hybridomas whose supernatants were found positive were
15 conserved and amplified before being tested by FACScan analysis so as to verify that the antibodies produced were likewise capable of recognizing native IGF-IR.

Example 2: Inhibition of [¹²⁵I]-IGF1 and [¹²⁵I]-IGF2 binding to human IGF-1 receptor

20 **Preparation of membrane lysate**

NIH 3T3 cells stably transfected with the human IGF-IR cDNA were grown in DMEM supplemented with 10 % of fetal calf serum. After detachment by scraping, serum-starved cells were further collected by centrifugation. The cell pellet was washed with phosphate buffered saline and resuspended in lysis buffer: 10 mM Tris-HCl pH 7.5
25 buffer containing protease inhibitors. Approximately 1 ml of buffer was added to 25.10⁶ cells. Cells were further lysed by 3 freeze-thawing cycles followed by 30 strokes of a Potter homogeniser at 1,900 rpm. After sonication, nuclei and large cell fragments were discarded by centrifugation at 1,000 g for 15 min at +4°C. Total cell membranes were obtained by centrifugation at 105,000 g for 1 h at +4°C. The membrane pellet was
30 washed in lysis buffer and centrifuged at 105,000 g for 1 h at +4°C. The final pellet was resuspended in 50 mM Tris-HCl buffer containing 150 mM NaCl, 0.5 % IGEPAL, 0.5 % Triton X-100, 0.25 % sodium deoxycholate and protease inhibitors, and stirred overnight at +4°C. Insoluble material was separated from the soluble extract containing

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hIGF-IR by centrifugation at 10,000 g for 10 min at +4°C. Membrane lysates were analyzed for protein concentration by the bicinchoninic assay and for IGF-IR by western blot.

[¹²⁵I]-IGF1 and [¹²⁵I]-IGF2 binding assays

5 The commercially available monoclonal antibody 17-69 (Neomarkers, Fremont, CA, USA), which has been described to recognize the IGF-IR alpha-subunit, was first coated on Protein A FlashPlate® 96-well microplates (Perkin Elmer, Boston, MA, USA) to immobilize IGF-IR. Two hundred µl of a 20 µg/ml antibody solution in PBS were added to each well and incubated overnight at +4°C. The buffer containing
10 residual 17-69 not attached to protein A was removed by aspiration. Two hundred µl of the membrane lysate at 100 µg/ml were further added and incubated for 2 h at room temperature to immobilize IGF-IR. Non captured proteins were removed by aspiration. For competition assays, binding of [¹²⁵I]-IGF1 (Perkin Elmer, Boston, MA, USA) or [¹²⁵I]-IGF2 (Amersham Biosciences, Saclay, France) at 100 pM to immobilized IGF-IR
15 was measured in the presence of varying concentrations of the monoclonal antibody I-3466 or the ligands IGF1, IGF2 and insulin (Sigma, Saint-Quentin Fallavier, France) ranging from 1 pM to 1 µM in binding buffer containing 50 mM Hepes pH 7.6, 150 mM NaCl, 0.05 % Tween 20, 1 % bovine serum albumin and 1 mM PMSF. The plates were incubated at room temperature for 2 h, then counted on a Packard Top Count Microplate
20 Scintillation Counter. Non specific binding was determined in the presence of 1 µM of IGF1. The monoclonal antibody 9G4, which is not directed at hIGF-IR but specifically recognizes an *E. coli* protein, was used as mouse IgG1 isotype control.

Results

25 Percent of specific [¹²⁵I]-IGF1 and [¹²⁵I]-IGF2 binding was plotted as a function of ligand concentration on semilog graphs. Concentrations of the various inhibitors required to inhibit the radioligand binding by 50 % (IC₅₀) were determined graphically from the sigmoid competition curves obtained (Figures 1 and 2).

30 Both IGF1 and IGF2 ligands efficiently displaced [¹²⁵I]-IGF1 binding to immobilized hIGF-IR, whereas insulin and the isotype control antibody 9G4 were unable to inhibit [¹²⁵I]-IGF1 binding at concentration lower than 500 nM (Figure 1). The monoclonal antibody I-3466 was able to inhibit the binding of [¹²⁵I]-IGF1 with an IC₅₀

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of 0.021 nM (Figure 1), which is 30-fold lower than the IC_{50} determined for non radiolabeled IGF1.

Moreover, the antibody I-3466 also exhibited a strong binding inhibition activity of [125 I]-IGF2 for immobilized hIGF-IR (Figure 2). In this case, the IC_{50} value deduced from the competition curve was around 0.1 nM. This IGF2 blocking activity of I-3466 was similar to the inhibitory activity induced by IGF2 ($IC_{50} = 0.06$ nM). This was slightly lower than the inhibitory activity of IGF1 ($IC_{50} = 0.02$ nM) and significantly greater than that of insulin ($IC_{50} \sim 200$ nM). As expected, the control antibody 9G4 did not show any IGF2 blocking activity.

10

Example 3: *In vitro* effect of the I-3466 antibody on either IGF1 or IGF2-induced MCF-7 growth

As indicated above, IGF-IR is overexpressed by numerous tumors but it has furthermore been described that in a significant number of breast and colon cancers, the proliferation signal is given to this receptor via IGF2 (sometimes written IGF-2, IGF-II or IGFII). It is therefore essential to ensure that the MAbs I-3466 is likewise capable of inhibiting both the IGF1 and the IGF2-induced growth on MCF-7 cells *in vitro*. In order to do this, cells were plated into 96-well plates at a density of 5×10^4 cells/well in 200 μ l of serum free medium (phenol red free-RPMI medium plus 1 % L-Glutamine). Twenty four hours after plating, either IGF1 at a final concentration of 50 μ g/ml (6.6 nM) or IGF-2 (final concentration of 100 ng/ml (13.2 nM)) was added to MCF-7 cells either in presence or in absence of I-3466 or a murin non neutralizing anti-IGF-IR antibody (7G3) used as an isotype control for nearly 52 additional hours.

Antibodies were tested at final concentrations ranging from 10 μ g/ml (66 nM) and 0.0097 μ g/ml (0.065 nM). Then cells were pulsed with 0.25 μ Ci of [3 H]thymidine for 16 hours and the magnitude of [3 H]thymidine incorporated in DNA was quantified by liquid scintillation counting. Both IGF-1 and IGF-2 significantly stimulate MCF-7 cell growth (Table 2).

No significant inhibition was observed when cells were treated with increasing doses of the isotype control antibody.

In contrast, when cells were incubated with increasing doses of the I-3466 antibody a significant dose dependant inhibition of both IGF1 (90 %) and IGF2 (84 %)-

30

induced proliferation was observed with IC₅₀s of 0.7 nM and 0.5 nM respectively (Table 2 and Figures 3A-3B).

Table 2

5 *In vitro* effect of the I-3466 antibody on either IGF1 (A)- or IGF2 (B)-induced MCF-7 growth

A		B	
	cpm		cpm
Cells alone	1449	Cells alone	1877
IGF1 50 ng/ml	67356	IGF2 100 ng/ml	39541
7G3 0,0097 µg/ml + IGF1	72144	7G3 0,0097 µg/ml + IGF2	60902
7G3 0,0195 µg/ml + IGF1	63200	7G3 0,0195 µg/ml + IGF2	67617
7G3 0,039 µg/ml + IGF1	53391	7G3 0,039 µg/ml + IGF2	56688
7G3 0,078 µg/ml + IGF1	56523	7G3 0,078 µg/ml + IGF2	60787
7G3 0,156 µg/ml + IGF1	46147	7G3 0,156 µg/ml + IGF2	49111
7G3 0,312 µg/ml + IGF1	54582	7G3 0,312 µg/ml + IGF2	48619
7G3 0,625 µg/ml + IGF1	53000	7G3 0,625 µg/ml + IGF2	51277
7G3 1,25 µg/ml + IGF1	58341	7G3 1,25 µg/ml + IGF2	54369
7G3 2,5 µg/ml + IGF1	51107	7G3 2,5 µg/ml + IGF2	53564
7G3 5 µg/ml + IGF1	45938	7G3 5 µg/ml + IGF2	54188
7G3 10 µg/ml + IGF1	34391	7G3 10 µg/ml + IGF2	55125
I-3466 0,0097 µg/ml + IGF1	63047	I-3466 0,0097 µg/ml + IGF2	45066
I-3466 0,0195 µg/ml + IGF1	56376	I-3466 0,0195 µg/ml + IGF2	43890
I-3466 0,039 µg/ml + IGF1	48403	I-3466 0,039 µg/ml + IGF2	31063
I-3466 0,078 µg/ml + IGF1	38986	I-3466 0,078 µg/ml + IGF2	27852
I-3466 0,156 µg/ml + IGF1	33595	I-3466 0,156 µg/ml + IGF2	18115
I-3466 0,312 µg/ml + IGF1	27966	I-3466 0,312 µg/ml + IGF2	13708
I-3466 0,625 µg/ml + IGF1	17166	I-3466 0,625 µg/ml + IGF2	9400
I-3466 1,25 µg/ml + IGF1	10791	I-3466 1,25 µg/ml + IGF2	7465
I-3466 2,5 µg/ml + IGF1	8281	I-3466 2,5 µg/ml + IGF2	6210
I-3466 5 µg/ml + IGF1	5721	I-3466 5 µg/ml + IGF2	6128
I-3466 10 µg/ml + IGF1	6823	I-3466 10 µg/ml + IGF2	7427

10 **Example 4: I-3466 inhibit both the IGF1- and IGF2-induced phosphorylation of the IGF-IR β-chain**

1.5 Either MCF7 or HT29 cells were cultured for 24 hours at 5.10⁴ cells/cm² (75 cm² plates, COSTAR) in 20 ml of RPMI without phenol red, mixed with 5 mM of glutamine, penicillin/streptomycin (respectively 100 U/100 µg/ml) and 10 % of fetal calf serum. After three washes in PBS, cells were incubated for 12 hours in phenol red-free medium (RPMI) without fetal calf serum and mixed with 5 mM of glutamine, penicillin/streptomycin, bovine serum albumin at 0.5 µg/ml (Sigma A-8022) and

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transferrin at 5 µg/ml (Sigma T8158).

For activation, cells were first incubated at 37°C for 15 minutes with blocking antibodies (10 µg/ml) to be tested and then either IGF1 or IGF2 was added for two additional minutes. The reaction was stopped by aspiration of the incubation medium and the plates were laid on ice. Cells were solubilized by addition of 0.5 ml of lysis buffer (50 mM tris-HCl pH 7.5, 150 mM NaCl, 1 % Nonidet P40, 0.5 % sodium deoxycholate), mixed with protease inhibitors (1 tablet per 50 ml, Boehringer Ref.: 1697 498), and phosphatase inhibitors (Calbiochem Ref.: 524625 (1/100)). Cells were scraped off and the suspension was recovered and placed on a shaker at 4°C for 1.5 hours. The solutions were centrifuged at 12,000 rpm for ten minutes (4°C) and the protein concentrations of the supernatants were quantified by BCA.

500 µg of proteins of the cell lysate were mixed with the anti-IGF-IR (Santa cruz Ref.: sc-713) for immunoprecipitation and incubated on the shaker at 4°C for 1.5 hours. The immunoprecipitates were recovered by addition of protein A-agarose (Boehringer Ref.: 1 134 515) and incubated all night on the shaker at 4°C. The agarose beads were washed twice with 1 ml of lysis buffer, twice with a wash buffer 1 (50 mM tris-HCl pH 7.5; 500 mM NaCl; 0.1 % Nonidet P40; 0.05 % sodium deoxycholate (Boehringer 1 332 597), mixed with protease inhibitors and phosphatase inhibitors) and once with a wash buffer 2 (50 mM tris-HCl; 0.1 % Nonidet P40; 0.05 % sodium deoxycholate (Boehringer Ref.: 1 332 597), mixed with protease inhibitors and phosphatase inhibitors 1/100). The immunoprecipitates were resuspended in a Laemmli buffer, heated to 100°C for 5 minutes. The supernatants were analyzed by electrophoresis on polyacrylamide SDS gel (8 % Novex EC6015). The proteins were transferred to a nitrocellulose membrane followed by either an immunoblot with anti-phosphotyrosine antibodies conjugated to HRP (BD transduction Labs PY20) or beta anti-chain of IGF-IR (Santa Cruz Ref.: sc 713) followed by an anti-rabbit antibody conjugated to HRP. The imprints were revealed by chemiluminescence (Amersham RPN 2209) followed by autoradiography on Kodak X-mat AR films.

Figures 4A and 4B represent MCF-7 cells nonstimulated (lanes 1, panel A and B) or stimulated either with 50 ng/ml IGF1 alone (lane 2, panel A) or 100 ng/ml IGF2 (lanes 2, panel B). As expected no basal stimulation of IGF-IR was observed in MCF-7 cells while a significant phosphorylation of the IGF-IR β-chain was observed when

MCF-7 cells were incubated either with IGF1 or IGF2. No stimulation was observed when cells were treated with an IgG1 isotype control (lanes 3, panel A and B) or with the I-3466 antibody alone (lanes 4, panel A and B) demonstrating that I-3466 displayed no agonistic effect on the IGF-IR. When I-3466 was added either with IGF1 or IGF2, a complete inhibition of the ligand-induced phosphorylation was observed (lanes 5, panel A and B). The 9G4 antibody used as isotype control was without any effect on either IGF1 or IGF2-induced phosphorylation (lanes 6, panel A and B).

The same inhibitory activity of I-3466 was observed on HT29 cells stimulated either with IGF1 or IGF2 (Figures 5A and 5B). These results are in agreement with the ones described in example 2 showing that I-3466 is able to displace both IGF1 and IGF2 from IGF-IR.

EXAMPLE 5: Internalization and degradation studies of the IGF-IR

Internalization and degradation studies were analyzed by FACS analysis. Internalization studies were performed using a murine biotinylated anti-IGF-IR monoclonal antibody (Mab) thereafter described as 12B1 Mab and binding to an epitope different from the one recognized by I-3466 antibody. The 9G4 Mab, was introduced as an isotype control. Both antibodies were generated in our laboratory. Confluent MCF-7 cells were trypsinized and 1×10^6 cells from each cellular suspension was plated in 96-well plates in FACS buffer. Plates were incubated, 4 hours at 37°C with either IGF1 (50 ng/ml) or with 30 µg/ml of I-3466, 9G4, mIgG1.

Cells incubated with FACS buffer alone were used to determine the basal level of expression of the IGF-IR.

Then cells were washed twice and 20 µg/ml of biotinylated-12B1 MAb were added to the plate. After 30 min of incubation at 4°C to avoid receptor internalization, cells were washed 3 times at 4°C and stained by addition of a streptavidin Alexa Fluor® 488-conjugate (Molecular Probes Europe BV, Leiden, Netherlands). For degradation experiments an additional step of cell permeabilization was added before staining cells with biotinylated-12B1 and streptavidin Alexa Fluor® 488-conjugate.

Table 3 showed that I-3466 causes a down regulation of the IGF-IR on both MCF-7 and HT29 cells after a 4 hours incubation period. No down regulation was observed when cells were incubated with the 9G4 Mab, used as an isotype control.

Table 3

Internalization/degradation experiments of IGF-IR by the I-3466 antibody

A- Internalization experiment

cell line ID	Tested Mab	MFI 12B1-mIgG1	% internalization
MCF-7	PBS	247	86
	9G4	226	
	I-3466	31	
HT29	PBS	69	33
	9G4	63	
	I-3466	42	

B- Degradation experiment

cell line ID	Tested Mab	MFI 12B1-mIgG1	% degradation
MCF-7	PBS	112	65
	9G4	105	
	I-3466	37	
HT29	PBS	34	52
	9G4	48	
	I-3466	23	

5

Example 6: Anti-tumor effects of I-3466 Mab on DU145, SK-ES-1, HT29, A549 and MCF-7 xenograft tumor models

To explore the activity of I-3466 antibody on *in vivo* tumor growth, five xenograft tumor models have been used: the androgen independent DU145 prostate cancer, the osteosarcoma SK-ES-1, the colon cancer HT29, the non-small cell lung cancer A549 and the breast cancer MCF-7. For that purpose, female athymic 6-8-week-old nude mice were injected subcutaneously with $2 \cdot 10^6$, $5 \cdot 10^7$, $5 \cdot 10^6$, $10 \cdot 10^6$ and $5 \cdot 10^6$ for DU-145, SK-ES-1, HT29, A549 and MCF-7 respectively. For DU145 and SK-ES-1 mice were treated 24 hours after cell injection with 200 μ g of non purified antibody.

Treatment was repeated twice a week. Concerning HT29, the treatment started when tumours reach a volume comprised between 49 and 59 mm³ and mice were injected i.p., 3 times a week, with 0.5 mg of purified antibody.

For A549, initial tumor volumes were comprised between 38 to 43 mm³ and for the MCF-7 experiments, they were comprised between 42 and 59 mm³ at the beginning of the treatment. Tumor volume was evaluated once or twice a week and calculated by

20

the following formula: $\pi/6 \times \text{length} \times \text{width} \times \text{height}$.

Figures 6A-6E showed results partly performed with non purified antibody indicating that I-3466 is able to significantly inhibit *in vivo* tumour growth of the 5 tested cell lines.

5

Example 7: Cloning, production and characterization of the humanized I-3466 IgG1 antibody

Chemically synthesized variable regions for light and heavy chains of the humanized antibody I-3466 were PCR amplified and cloned into antibody expression
10 vectors carrying human light chain kappa and heavy chain IgG1 constant regions, respectively. The PCR primers contain 36 nucleotides with 15 nucleotides for In-Fusion annealing with the vector overlapping cloning region and 21 nucleotides for annealing with the variable regions. The exact nucleotide sequences are shown below:

for light chain variable region cloning:

15 IF-Humanized I-3466-LCK-F (SEQ ID No. 21)
[5'-ACAGATGCCAGATGCGACATTGTGATGACCCAGTCC],
IF-Humanized I-3466-LCK-R (SEQ ID No. 22)
[5'-TGCAGCCACCGTACGCTTGATCTCCACCTTGGTGCC],

for heavy chain variable region cloning:

20 IF-Humanized I-3466-HCG1-F (SEQ ID No. 23)
[5'-ACAGGTGTCCACTCGGAGGTGCAGCTGGTGGAGTCT],
IF-Humanized I-3466-HCG1-R (SEQ ID No. 24)
[5'-GCCCTTGGTGGATGCGGAGGAGACTGTCACCAGGGT].

The vectors were linearized with BmtI and FspI digestion. The In-Fusion
25 reaction was carried out according to vendor's recommended protocol. The resulting clones were confirmed by sequencing. Human embryonic kidney 293 cells were transfected using light and heavy chain plasmid DNAs. Medium containing secreted antibody was harvested and concentrated. The antibody was affinity purified using protein A/G column and concentrated and dialyzed in PBS. The protein concentration
30 was determined by OD280nm and the purity was analyzed by reducing and non-reducing SDS-PAGE (Figure 9).

Example 8: BIAcore Analysis of Humanized I-3466 IgG1 antibody**Equipment and materials**

BIAcore T100 instrument, CM5 biosensor chips, HBS-EP buffer, acetate buffer (pH 5), Glycine-HCl buffer (pH 1.5), amine coupling kit were from BIAcore (Uppsala, Sweden). Anti-human IgG Fc was from Jackson ImmunoResearch Laboratories Inc. (West Grove, USA), soluble human insulin-like growth factor-1 receptor (hIGF-IR) extra-cellular domain (ECD) was from R&D Systems (Minneapolis, USA).

Biacore Assay

All experiments were performed at 25°C at a flow rate of 40 µl/min. To prepare a BIAcore assay (see Figure 10), an anti-human IgG-Fc antibody (50 µg/ml in acetate buffer, pH 5) was immobilized onto a carboxymethyl dextran sensorchip (CM5) using amine coupling procedures as described by the manufacturer. 11042 and 11111 resonance units (RU) of anti-IgG Fc antibodies were linked respectively onto Flowcells (FC) 1 and 3. Purified Mabs to be tested were diluted at a concentration of 5 µg/ml in 0.5 % P20, HBS-EP buffer and injected on FC3 to reach 500 to 1000 RU. FC1 was used as the reference cell. Specific signals correspond to the difference of signals obtained on FC2 *versus* FC1. The analyte (soluble hIGF-IR, apparent molecular weight 365 kDa) was injected during 90 sec at 5 different concentrations (100, 50, 25, 12.5 and 6.25 nM) in 0.5 % P20, HBS-EP buffer. These concentrations were prepared from stock solution in 0.5 % P20, HBS-EP. The dissociation phase of the analyte was monitored over a 10 minutes period. Running buffer was also injected under the same conditions as a double reference. After each cycle (antibody + hIGF-IR injection), both Flowcells were regenerated by injecting 20 to 45 µl of Glycine-HCl buffer (pH 1.5). This regeneration is sufficient to eliminate all Mabs and Mabs/hIGF-IR complexes captured on the sensorchip.

Result

The binding of the Mab Humanized I-3466 to the analyte hIGF-IR-ECD was characterized by the association and dissociation rate constants k_a and k_d , respectively. The equilibrium dissociation constant (KD) was calculated by the ratio between dissociation and association rate constants. Results are given in the following Table 4. The sensorgram corresponding to the different analyte concentrations is represented in the Figure 11.

Table 4

IGF-IR Abs	ka (1/Ms)	kd (1/s)	KD (M)
IgG1 HI3466	1.84E+05	6.29E-05	3.42E-10

CLAIMS

1. An isolated antibody, or a functional immunogenic fragment thereof, having a binding affinity for the human insulin-like growth factor I receptor (IGF-IR),
5 characterized in that upon binding said IGF-IR, it inhibits the binding of the native binding partner IGF1 to said IGF-IR with an IC50 of less than 0.3 nM, preferentially less than 0.03 nM and it also inhibits the binding of the native binding partner IGF2 to said IGF-IR with an IC50 of less than 0.3 nM, preferentially less than 0.1 nM.

2. An isolated antibody, or a functional immunogenic fragment thereof, having
10 a tyrosine kinase of the human insulin-like growth factor I receptor (IGF-IR) inhibiting activity, characterized in that upon binding said IGF-IR, it inhibits the binding of the native binding partner IGF1 to said IGF-IR with an IC50 of less than 0.3 nM, preferentially less than 0.03 nM and it also inhibits the binding of the native binding partner IGF2 to said IGF-IR with with an IC50 of less than 0.3 nM, preferentially less
15 than 0.1 nM.

3. The antibody, or one of its functional fragments, according to claim 1 or 2, characterized in that it is also capable of inhibiting 100 % of the IGF1 and/or IGF2 induced phosphorylation of the IGF-IR beta-chain.

4. The antibody, or one of its functional fragments, according to claims 1 to 3,
20 characterized in that it does not present any agonistic intrinsic activity.

5. The antibody, or one of its functional fragments, according to claims 1 to 4, characterized in that it is capable of inducing, using the same FACS analysis method:

- i) at least 30 % of the IGF-IR internalization on HT29 cells, and/or
- ii) at least 85 % of the IGF-IR internalization on MCF-7 cells.

6. The antibody, or one of its functional fragments, according to claims 1 to 5,
25 characterized in that it is capable of inducing, using the same FACS analysis method:

- i) at least 50 % of the IGF-IR degradation on HT29 cells, and/or
- ii) at least 65 % of the IGF-IR degradation on MCF-7 cells.

7. The antibody, or one of its functional fragments, according to claims 1 to 6,
30 characterized in that it is able to inhibit both IGF1- and IGF2-induced in vitro proliferation of MCF-7 cells with IC50s at least equal to 1 nM and preferentially at least equal to 0.7 and 0.5 nM respectively for IGF1 and IGF2.

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8. The antibody, or one of its functional fragments, as claimed in claims 1 to 7, characterized in that it comprises a light chain comprising at least one complementarity determining region CDR chosen from the CDRs of sequence SEQ ID No. 1, 3 or 5, or at least one CDR whose sequence has at least 80 % identity after optimum alignment with the sequence SEQ ID No. 1, 3 or 5, or in that it comprises a heavy chain comprising at least one CDR chosen from the CDRs of sequence SEQ ID No. 2, 4 or 6, or at least one CDR whose sequence has at least 80% identity after optimum alignment with the sequence SEQ ID No. 2, 4 or 6.

9. The antibody, or one of its functional fragments, as claimed in claims 1 to 8, characterized in that it comprises a heavy chain comprising at least two of the three CDRs or the three CDRs of sequence SEQ ID Nos. 2, 4 and 6, or at least two of three CDRs or three CDRs of sequence respectively having at least 80 % identity after optimum alignment with the sequence SEQ ID Nos. 2, 4 and 6.

10. The antibody, or one of its functional fragments, as claimed in one of claims 1 to 9, characterized in that it comprises a light chain comprising at least two of the three CDRs or the three CDRs of sequence SEQ ID Nos. 1, 3 and 5, or at least two of three CDRs or three CDRs of sequence respectively having at least 80 % identity after optimum alignment with the sequence SEQ ID Nos. 1, 3 and 5.

11. The antibody, or one of its functional fragments, as claimed in one of claims 1 to 10, characterized in that it comprises a heavy chain comprising the three CDRs of sequence SEQ ID Nos. 2, 4 and 6, or three CDRs of sequence respectively having at least 80 % of identity after optimum alignment with the sequence SEQ ID Nos. 2, 4 and 6 and in that it moreover comprises a light chain comprising the three CDRs of sequence SEQ ID Nos. 1, 3 and 5, or three CDRs of sequence respectively having at least 80 % of identity after optimum alignment with the sequence SEQ ID Nos. 1, 3 and 5.

12. The antibody, or one of its functional fragments, as claimed in one of claims 1 to 11, characterized in that it does not attach in a significant manner to the human insulin receptor (IR).

13. The antibody as claimed in one of claims 1 to 12, characterized in that said functional fragment is chosen from the fragments Fv, Fab, F(ab')₂, Fab', scFv, scFv-Fc and the diabodies, or any fragment whose half-life would have been increased such as

pegylated fragments.

14. A murine hybridoma capable of secreting an antibody as claimed in one of claims 1 to 13.

5 15. The murine hybridoma as claimed in claim 14 deposited at the CNCM, Institut Pasteur, Paris, on 23 June 2005 under the number I-3466.

16. An antibody, or one of its functional fragments, characterized in that said antibody is secreted by the hybridoma as claimed in claim 15.

10 17. The antibody, or one of its functional fragments, as claimed in one of claims 1 to 13 or 16, characterized in that said antibody comprises a light chain of sequence comprising the amino acid sequence SEQ ID No. 7, or a sequence having at least 80 % identity after optimum alignment with the sequence SEQ ID No. 7, or/and in that it comprises a heavy chain of sequence comprising the amino acid sequence SEQ ID No. 8, or a sequence having at least 80 % identity after optimum alignment with the sequence SEQ ID No. 8.

15 18. The antibody or one of its functional fragments, as claimed in claim 17, characterized in that said antibody is a chimeric antibody and moreover comprises the light chain and heavy chain constant regions derived from an antibody of a species heterologous to the mouse.

20 19. The chimeric antibody, or one of its functional fragments, as claimed in claim 18, characterized in that said heterologous species is man.

20. The chimeric antibody, or one of its functional fragments, as claimed in claims 17 to 19, characterized in that the light chain and heavy chain constant regions derived from a human antibody are respectively the kappa and gamma-1, gamma-2 or gamma-4 region.

25 21. The humanized antibody, or one of its functional fragments, as claimed in claim 19 or 20, characterized in that it comprises a light chain comprising the amino acid sequence SEQ ID No. 17, or a sequence having at least 80 % identity after optimum alignment with the sequence SEQ ID No. 17, or/and in that it comprises a heavy chain comprising the amino acid sequence SEQ ID No. 18, or a sequence having
30 at least 80 % identity after optimum alignment with the sequence SEQ ID No. 18.

22. The humanized antibody, or one of its functional fragments, as claimed in claim 21, characterized in that it comprises a light chain comprising the amino acid

sequence SEQ ID No. 17 and in that it comprises a heavy chain of sequence comprising the amino acid sequence SEQ ID No. 18.

23. An isolated nucleic acid, characterized in that it is chosen from the following nucleic acids:

- 5 a) a nucleic acid, DNA or RNA, coding for an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13 and 16 to 22;
- b) a complementary nucleic acid of a nucleic acid such as defined in a);
- c) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions of great stringency with at least one of the CDRs of sequence SEQ ID No. 9, 10, 11,
10 12, 13 or 14, or with a sequence having at least 80 % identity after optimum alignment with the sequence SEQ ID No. 9, 10, 11, 12, 13 or 14;
- d) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions of great stringency with at least the light chain of nucleic acid sequence SEQ ID No. 15 and/or the heavy chain of nucleic acid sequence SEQ ID No. 16, or with a
15 sequence having at least 80 %, preferably 85 %, 90 %, 95 % and 98 %, identity after optimum alignment with the sequence SEQ ID No. 15 and/or 16; and
- e) a nucleic acid of at least 18 nucleotides capable of hybridizing under conditions of great stringency with at least the light chain of nucleic acid sequence SEQ ID No. 19 and/or the heavy chain of nucleic acid sequence SEQ ID No. 20, or with a
20 sequence having at least 80 %, preferably 85 %, 90 %, 95 % and 98 %, identity after optimum alignment with the sequence SEQ ID No. 19 and/or 20.

24. A vector comprising a nucleic acid as claimed in claim 23.

25. A host cell comprising a vector as claimed in claim 24.

26. A transgenic animal with the exception of man comprising at least one cell
25 transformed by a vector as claimed in claim 25.

27. A process for production of an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13 and 16 to 22, characterized in that it comprises the following stages:

- 30 a) culture in a medium and appropriate culture conditions of a cell as claimed in claim 25; and
- b) recovery of said antibodies, or one of their functional fragments, thus produced starting from the culture medium or said cultured cells.

28. Antibody, or one of its functional fragments, capable of being obtained by a process as claimed in claim 27.

29. The antibody, or one of its functional fragments, as claimed in claims 1 to 13 and 16 to 22 and 28, characterized in that it is, moreover, capable of binding
5 specifically to a human receptor of the tyrosine kinase family and/or capable of inhibiting the tyrosine kinase activity of such receptors.

30. The antibody, or one of its functional fragments, as claimed in claim 29, characterized in that it consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the binding of EGF on the human epidermal growth
10 factor receptor (EGFR) and/or inhibiting the tyrosine kinase activity of said EGFR.

31. The antibody or one of its functional fragments, as claimed in claim 30, characterized in that said second anti-EGFR motif is issued from the murine monoclonal antibody 225, its chimeric derivative C225 or any humanized antibody derived from this antibody 225.

15 32. The antibody, or one of its functional fragments, as claimed in claim 29, characterized in that it consists in a bispecific antibody comprising a second motif capable of specifically inhibiting the activity modulated by the HER2/neu receptor and/or inhibiting the tyrosine kinase activity of said HER2/neu receptor.

33. The antibody or one of its functional fragments, as claimed in claim 32,
20 characterized in that said second anti-HER2/neu motif is issued from the murine monoclonal antibody 4D5 or 2C4 or the humanized antibody Trastuzumab or Pertizumab.

34. The antibody, or one of its functional fragments, as claimed in claim 29, characterized in that it consists in a bispecific antibody comprising a second motif
25 capable of specifically inhibiting the binding of the Hepatocyte Growth Factor (HGF) on the cMET receptor and/or inhibiting the tyrosine kinase activity of said cMET receptor.

35. The antibody, or one of its functional fragments, as claimed in claim 29, characterized in that it consists in a bispecific antibody comprising a second motif
30 capable of specifically inhibiting the binding of the Macrophage Stimulating Protein (MSP) on the RON receptor and/or inhibiting the tyrosine kinase activity of said RON receptor.

36. The antibody, or one of its functional fragments, as claimed in claims 1 to 13, 16 to 22 and 28 to 35, characterized in that it is capable to interact with any kind of receptor being implicated in the development of tumours such as, for example, VEGFR, FGF (Fibroblast Growth Factor) or CXCR4 or 2 (Chemokine receptor type 4 or 2).

5 37. The antibody as claimed in one of claims 28 to 36, characterized in that said second motif is selected from the fragments Fv, Fab, F(ab')₂, Fab', Fab'PEG, scFv, scFv-Fc and the diabodies, or any form whose half-life would have been increased.

38. The antibody, or one of its functional fragments, as claimed in one of claims 1 to 13, 16 to 22 and 28 to 37 as a medicament.

10 39. A composition comprising by way of active principle a compound consisting of an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13, 16 to 22 and 28 to 38.

40. A composition as claimed in claim 39, characterized in that it comprises at least a second compound chosen from the compounds capable of specifically inhibiting
15 the tyrosine kinase activity of IGF-IR, EGFR, HER2/neu, cMET and/or RON.

41. A composition according to claim 40, characterized in that said second compound is chosen from the isolated anti-EGFR, -IGF-IR, -HER2/neu, -cMET and/or -RON antibodies, or their functional fragments, capable of inhibiting the proliferative and/or anti-apoptotic and/or angiogenic and/or metastatic dissemination inductor
20 activity mediated by said receptor(s).

42. A composition as claimed in claims 39 to 41, characterized in that it comprises, as combination product for a simultaneous, separate or sequential use, at least an inhibitor of the tyrosine kinase activity of the IR, IGF-IR, EGFR, HER2/neu, cMET and/or RON.

25 43. A composition as claimed in claim 42, characterized in that said inhibitor of the tyrosine kinase activity is selected from the group consisting of derived natural agents, dianilinophthalimides, pyrazolo- or pyrrolopyridopyrimidines or else quinazolines.

44. A composition as claimed in claims 39 to 43, characterized in that it
30 comprises, moreover, as a combination product for simultaneous, separate or sequential use, a cytotoxic/cytostatic agent.

45. The composition as claimed in claim 44, characterized in that said cytotoxic/cytostatic agent is chosen from the agents interacting with DNA, the antimetabolites, the topoisomerase I or II inhibitors, or the spindle inhibitor or stabilizer agents or else any agent capable of being used in chemotherapy.

5 46. The composition as claimed in claim 44 or 45, characterized in that said cytotoxic/cytostatic agent is coupled chemically to at least one of the elements of said composition for simultaneous use.

10 47. The composition as claimed in claims 44 to 46, characterized in that said cytotoxic/cytostatic agent is chosen from the spindle inhibitor or stabilizer agents, preferably vinorelbine, vinflunine or vincristine.

48. The composition as claimed in any one of claims 39 to 47, characterized in that one, at least, of said antibodies, or one of its functional fragments, is conjugated with a cell toxin and/or a radioelement.

49. The composition as claimed in one of claims 39 to 48 as a medicament.

15 50. The use of an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13, 16 to 22 and 28 to 38 and/or of a composition as claimed in any one of claims 39 to 49 for the preparation of a medicament intended for the prevention or for the treatment of an illness connected with an overexpression and/or an abnormal activation of the IGF-IR, and/or connected with a hyperactivation of the transduction
20 pathway of the signal mediated by the interaction of IGF1 or IGF2 with IGF-IR.

51. The use as claimed in claim 50, characterized in that the administration of said medicament does not induce or only slightly induces secondary effects connected with inhibition of the insulin receptor IR.

25 52. The use as claimed in claim 50 or 51 for the preparation of a medicament intended to inhibit the transformation of normal cells into cells with tumoral character and/or the growth and/or the proliferation of tumor cells, preferably IGF-dependent, at least IGF2-dependent cells.

53. The use as claimed in one of claims of 50 to 52 for the preparation of a medicament intended for the prevention or for the treatment of cancer.

30 54. The use as claimed in claim 53, characterized in that said cancer is a cancer chosen from prostate cancer, osteosarcomas, lung cancer, breast cancer, endometrial cancer, multiple myeloma, ovary cancer or colon cancer.

55. The use as claimed in claim 54, characterized in that said cancer is colon cancer.

56. The use as claimed in one of claims 50 to 52 for the preparation of a medicament intended for the prevention or for the treatment of psoriasis or
5 atherosclerosis.

57. A method of in vitro diagnosis of pathological condition characterized by aberrant expression of IGF-IR relative to normal comprising contacting a biological sample suspected of containing IGF-IR, with the antibody as claimed in one of claims 1 to 13, 16 to 22 and 28 to 38, under conditions favoring formation of an IGF-IR/antibody
10 complex, and detecting said complex as indicating presence said IGF-IR in said sample.

58. The method of claim 57, wherein said antibody is detectably labeled.

59. The method according to claim 57 or 58, wherein the aberrant expression of IGF-IR is an over-expression of IGF-IR.

60. The method according to claim 57 or 58, wherein the aberrant expression of IGF-IR is an under-expression of IGF-IR.
15

61. A diagnostic method for predicting an oncogenic potential of a sample of prostate cells, comprising:

- (a) providing a sample of human lung or breast tissue; and
- 20 (b) determining, in the sample, presence of IGF-IR, said step comprising contacting said sample with the antibody according to one of claims 1 to 13, 16 to 22 or 28 to 38 under conditions favoring formation of an IGF-IR/antibody complex, wherein presence of said complex indicates the oncogenic potential of said cells in said tissue.

62. The method according to claim 61, wherein presence of said complex indicates that the patient is at risk of developing or onset of a pathological disorder characterized by overexpression of said IGF-IR.
25

63. A method for following progress of a therapeutic régime designed to alleviate a pathological disorder characterized by an abnormal expression of IGF-IR expression, comprising:
30

- (a) assaying a sample from a subject to determine level of IGF-IR at a first time point;
- (b) assaying said sample at a second time point; and

- (c) comparing said level at said second time point to the level determined in (a) as a determination of effect of said therapeutic regime wherein a decrease in the level of IGF-IR in said sample is determinative of the regression said pathological disorder in said patient or an increase in the level of IGF-IR is determinative of the progression of said pathological disorder in said patient.

64. A kit or set for carrying out a method of diagnosis of illnesses induced by an overexpression or an underexpression of the IGF-IR or for carrying out a process for the detection and/or the quantification of an overexpression or of an underexpression of the IGF-IR in a biological sample, preferably an overexpression of said receptor, characterized in that said kit or set comprises the following elements:

- a) an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13, 16 to 22 and 28 to 38;
- b) optionally, the reagents for the formation of the medium favorable to the immunological reaction;
- c) optionally, the reagents allowing the demonstration of IGF-IR/antibody complexes produced by the immunological reaction.

65. The use of an antibody, or one of its functional fragments, as claimed in one of claims 1 to 13, 16 to 22 and 28 to 38, for the preparation of a medicament intended for the specific targeting of a biologically active compound to cells expressing or overexpressing the IGF-IR.

66. A method of modulating IGF-IR activity in IGF-IR-responsive mammalian cells comprising: contacting the cells with an antibody as claimed in claims 1 to 13, 16 to 22 and 28 to 38 under conditions sufficient to modulate activity of said IGF-IR relative to non-IGF-IR responsive cells.

67. A method of decreasing IGF-IR activity in IGF-IR-responsive mammalian cells relative to normal cells comprising: contacting the cells with an antibody as claimed in claims 1 to 13, 16 to 22 and 28 to 38.

68. A method of identifying an IGF-IR modulator comprising:

- a) contacting IGF-IR with the antibody as claimed in claims 1 to 13, 16 to 22 and 28 to 38;
- b) contacting the complex of (a) with a compound library;
- c) identifying a compound which disrupts the complex of (a); and

- 62 -

d) determining whether the compound exhibits agonist or antagonist activity at the IGF-IR, wherein this activity indicates identification of an IGF-IR modulator.

69. A method of detecting a binding partner for the antibody of claims 1 to 13, 16 to 22 or 28 to 38 in a sample, the method comprising: a) incubating the antibody of
5 claims 1 to 13, 16 to 22 or 28 to 38 with a biological sample obtained from a patient presenting with a cell proliferative disorder characterized by abnormal level of expression and/or activation of human IGF-IR under conditions sufficient to allow specific binding of the antibody to its binding partner and b) detecting specific binding, wherein specific binding indicates the presence of a binding partner in the sample.

10 70. A method of purifying a IGF-IR from a sample, the method comprising: a) incubating the antibody of claim 1 to 13, 16 to 22 or 28 to 38 with a sample under conditions to allow specific binding of the antibody and the receptor, and b) separating the antibody from the sample and obtaining the purified receptor.

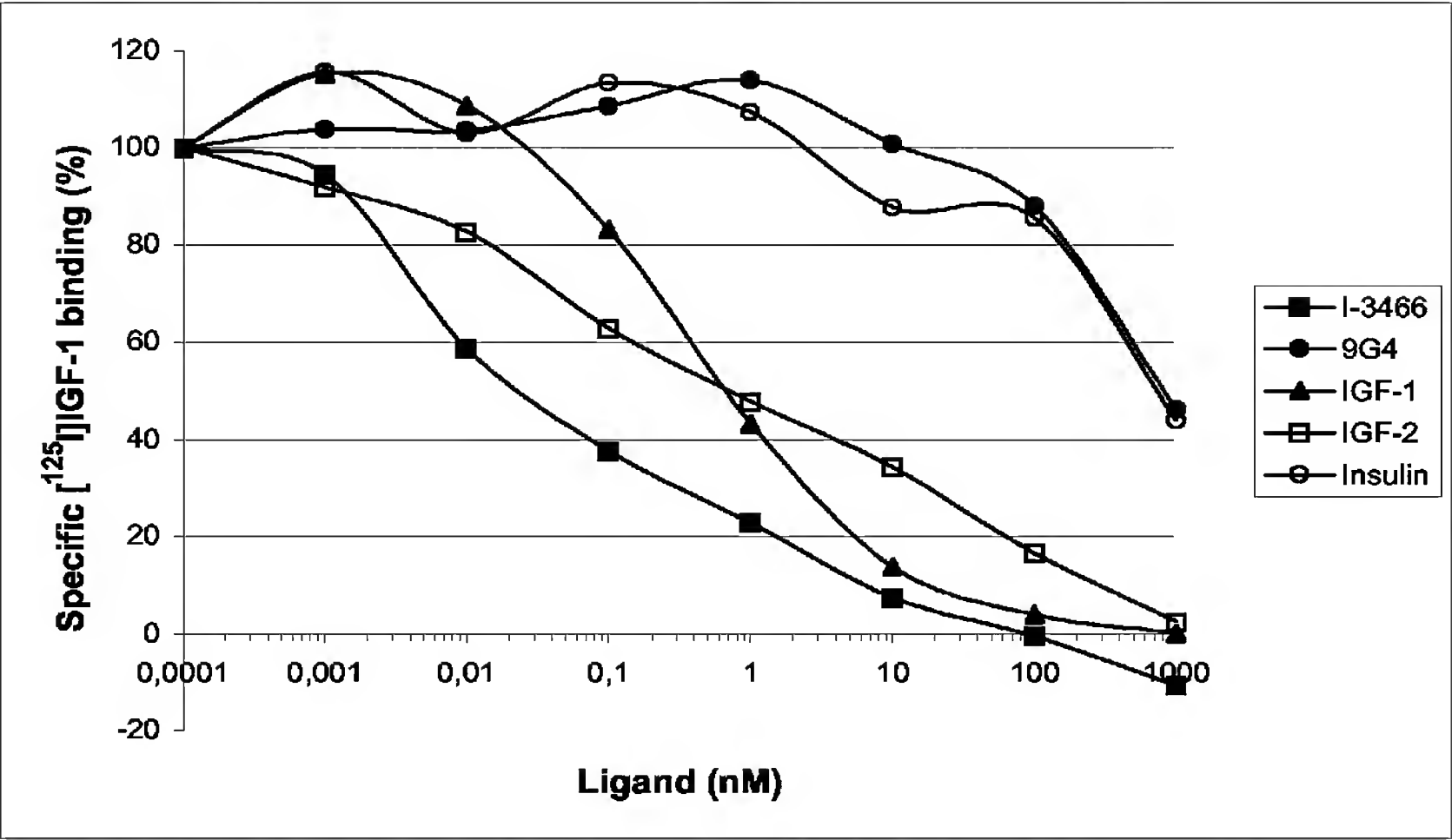


FIGURE 1

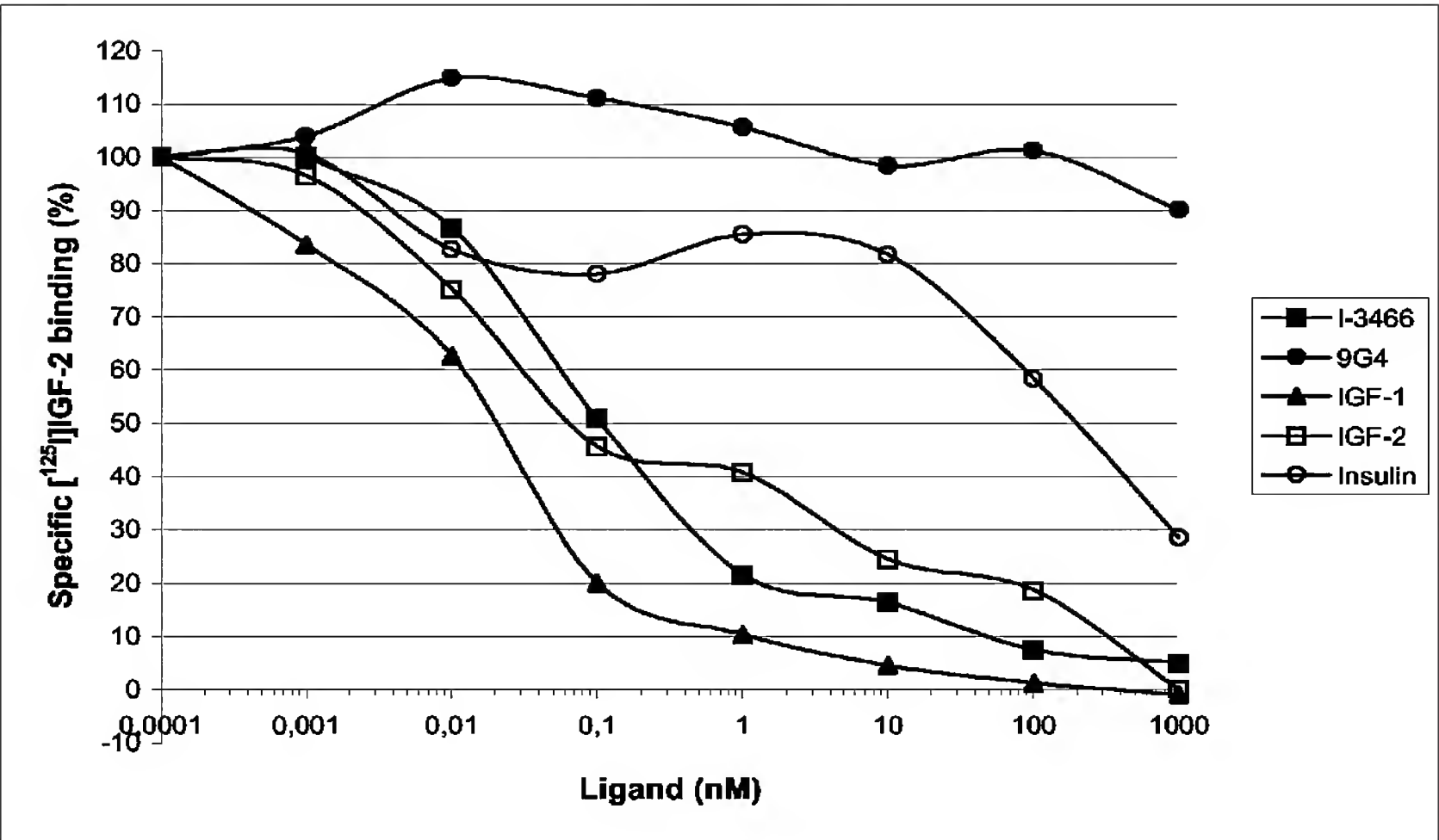
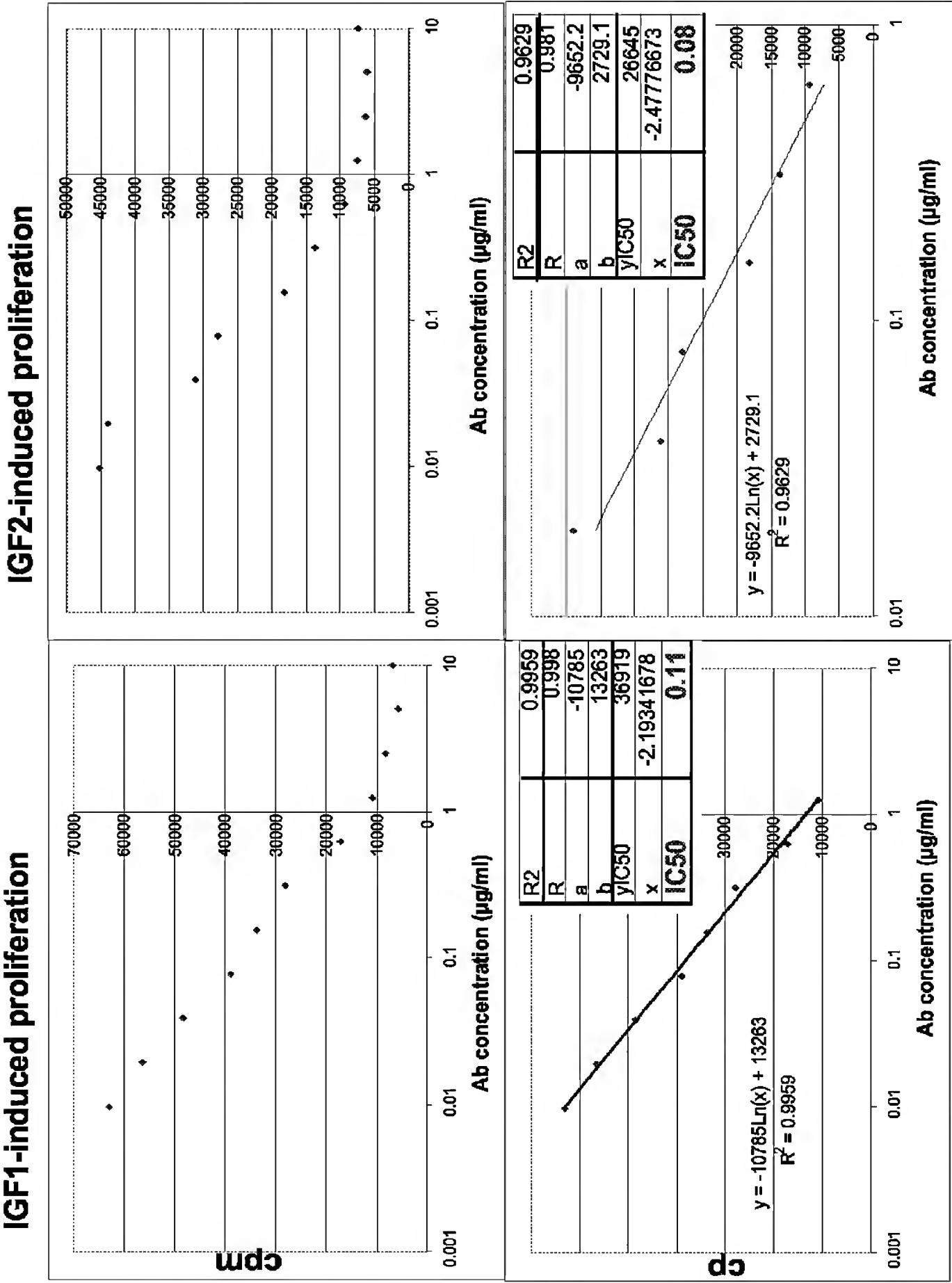


FIGURE 2



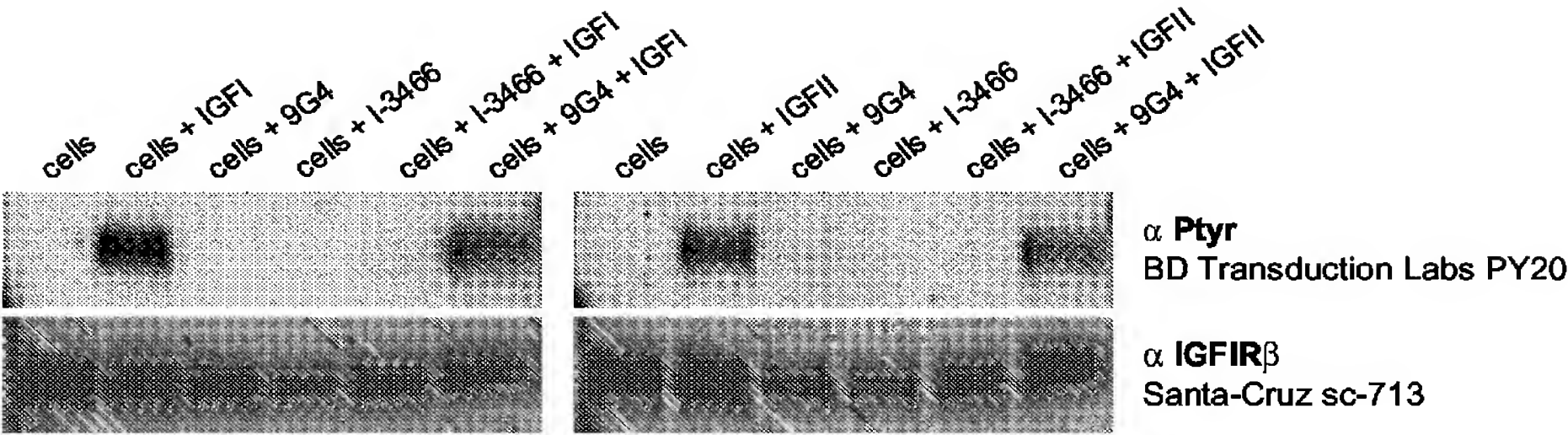


FIGURE 4A

FIGURE 4B

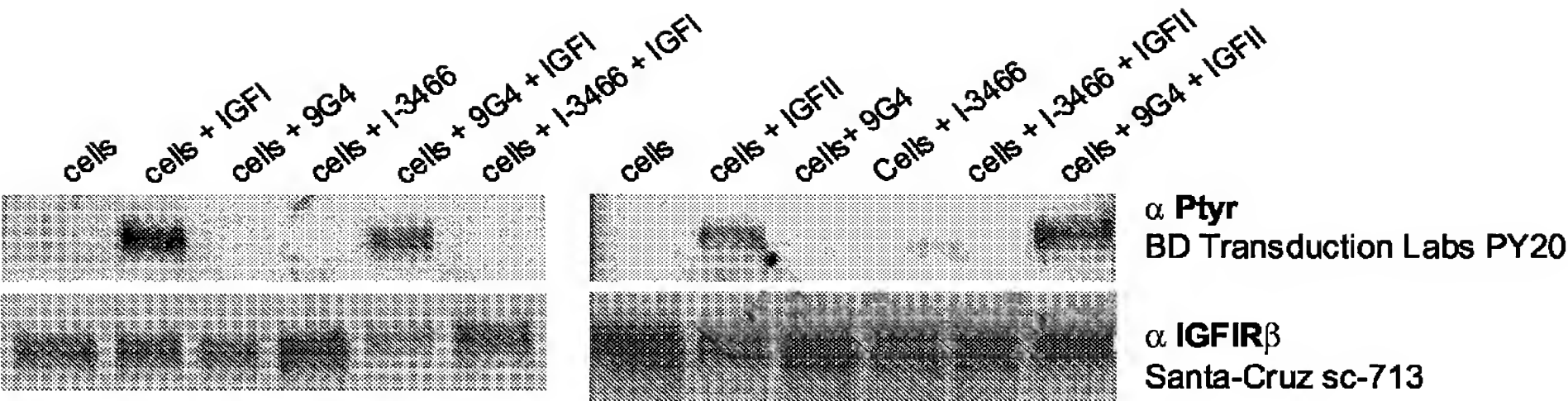


FIGURE 5A

FIGURE 5B

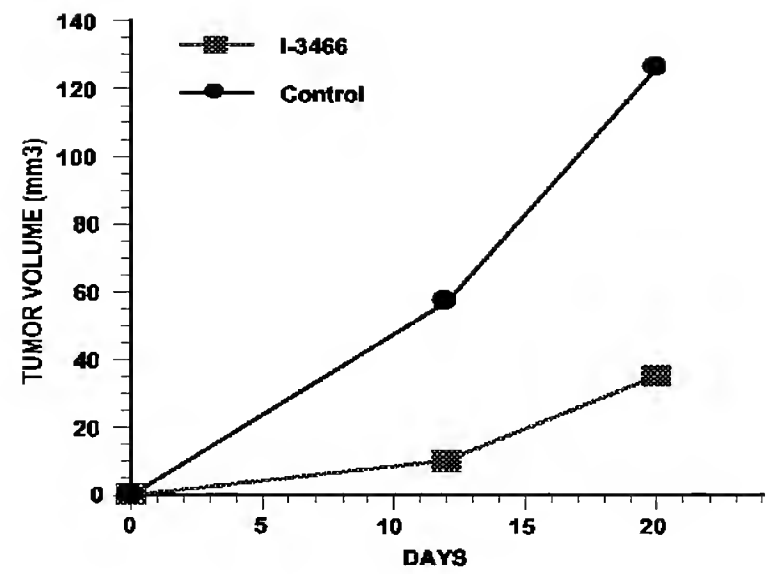


FIGURE 6A

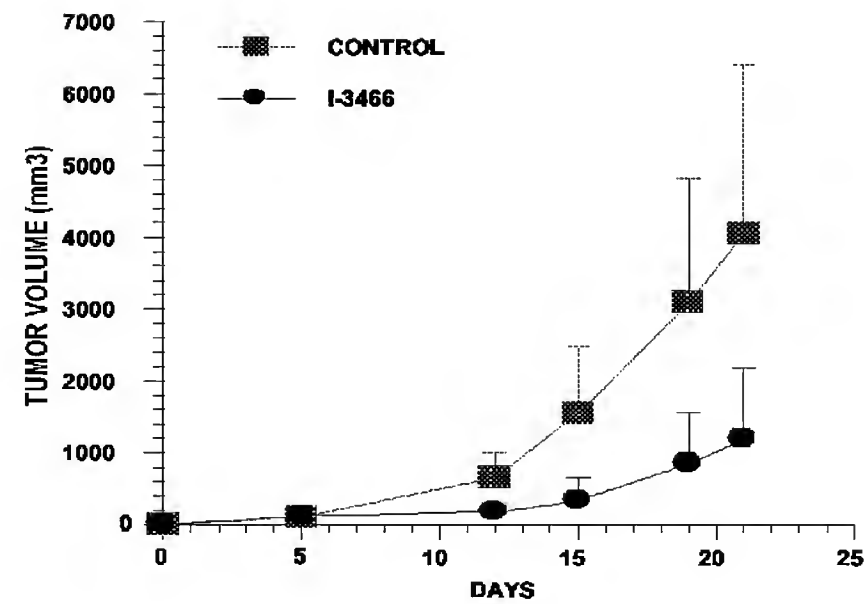


FIGURE 6B

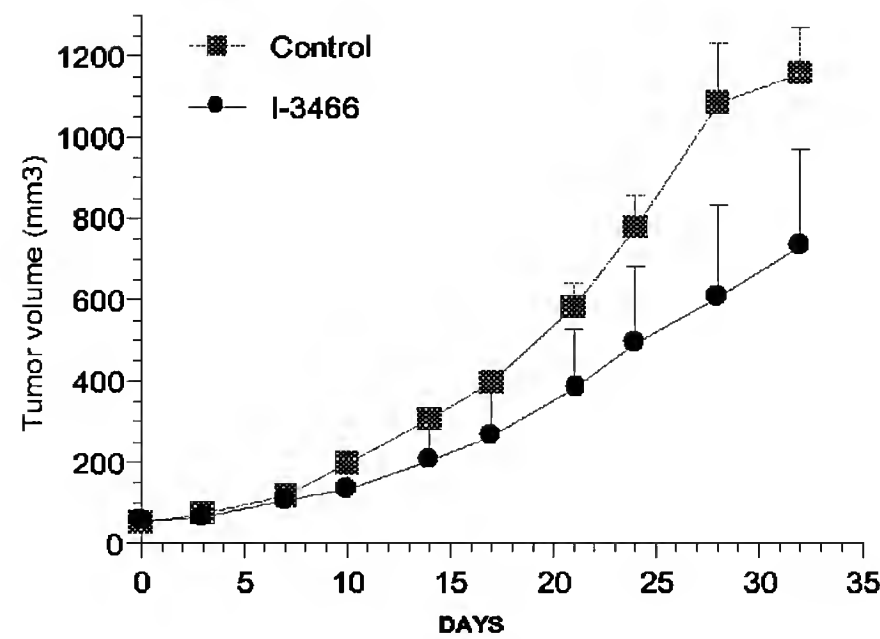


FIGURE 6C

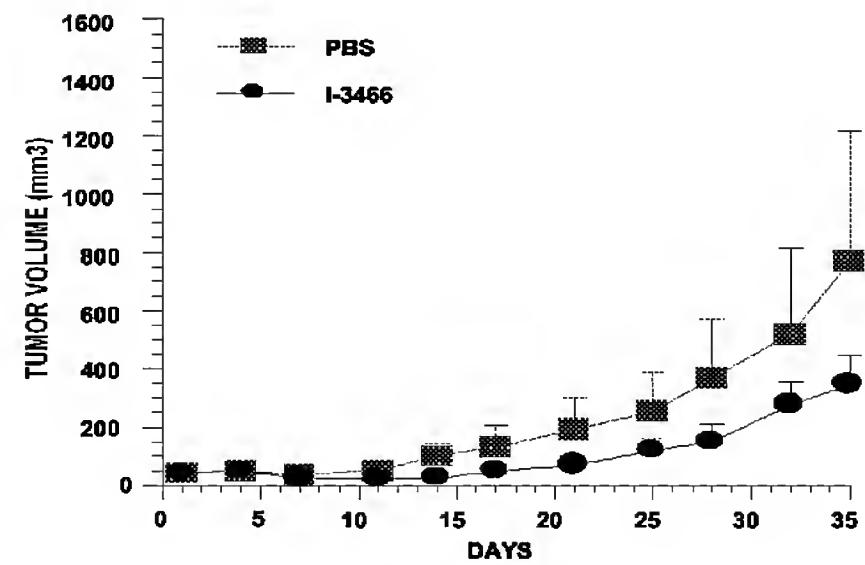


FIGURE 6D

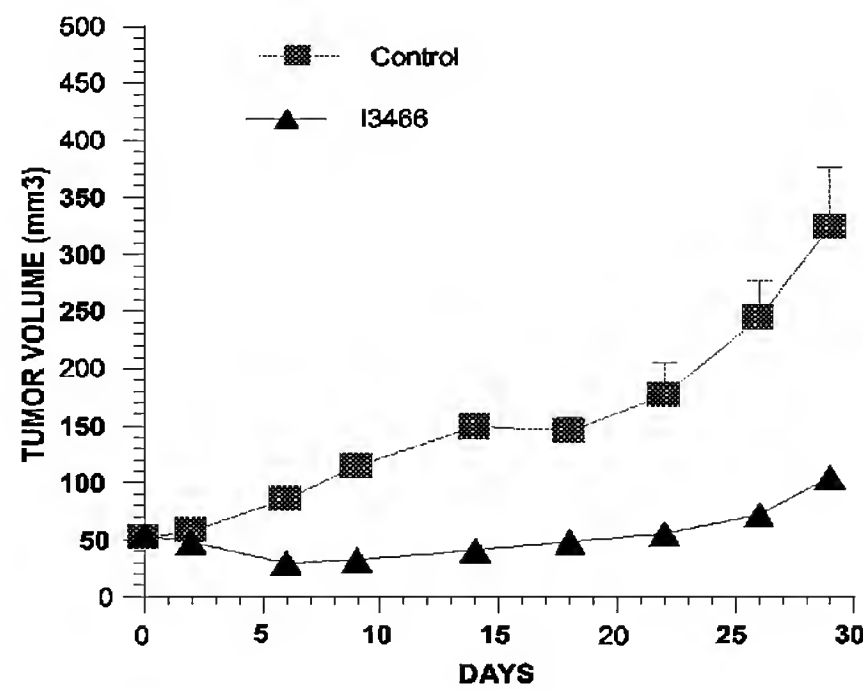


FIGURE 6E

5/7

	1*		* *		*# # # #
mouse I-3466 LCVR	DIVMSQSPSS	LAVSAGEKVT	MNCKSSQSLL	DSRTRKNYLA	WYQQKPGQSP
P06313	DIVMTQSPDS	LAVSLGERAT	INCKSSQSVL	YSSNNKNYLA	WYQQKPGQPP
VKII 3-1-(1) 011	DIVMTQTPLS	LPVTFGEFAS	ISCRSSQSLL	DSDDGNTYLD	WYLQKPGQSP
Humanized I-3466 LCVR	DIVMTQSPDS	LAVSLGERAT	INCKSSQSLL	DSRTRKNYLA	WYQQKPGQPP

	# * **	*	*		# #*#
mouse I-3466 LCVR	KLLIYWASTR	ESGVPDRFTG	SGSGTDFTLT	ISSVQAEDLA	VYYCKQSYNL
P06313	KLLIYWASTR	ESGVPDRFSG	SGSGTDFTLT	ISSLQAEDVA	VYYCQYDTI
VKII 3-1-(1) 011	QLLIYTLSYR	ASGVPDRFSG	SGSGTDFTLK	ISRVEAEDVG	VYYCNQRIEF
Humanized I-3466 LCVR	KLLIYWASTR	ESGVPDRFSG	SGSGTDFTLT	ISSLQAEDVA	VYYCKQSYNL

	*##	113
mouse I-3466 LCVR	FTFGGGTKLE	IK-
P06313	FTFGGGTKVE	IKR
VKII 3-1-(1) 011	F	
Humanized I-3466 LCVR	FTFGGGTKVE	IKR

FIGURE 7

	1		* ** *		*# # #		# # 50
mouse I-3466 HCVR	EVMLVESGGD	LVKPGGSLKL	SCAASGFTFN	NYIMSWVRQT	PEKRLWVAT		
BAC02119	EVQLVESGGD	LVQPGGSLRL	SCAASGFTFT	NYAMSWVRQA	PGKGLEWVSA		
VH3 1-3 3-30	QVQLVESGGG	VVQPGRLRL	SCAASGFTFS	SYGMWVRQA	PGKGLEWVAV		
Humanized I-3466 HCVR	EVQLVESGGD	LVQPGGSLRL	SCAASGFTFN	NYIMSWVRQA	PGKGLEWVAT		

	51	*	*		# #*#
mouse I-3466 HCVR	ISGGGSYTFY	PDSVKGRFSI	SRDNAKNTLY	LQMSSLRSED	TAMYFCTRNO
BAC02119	ISGGGSYTFY	ADSVKGRFTI	SRDNSKNTLY	LQMNSLRAED	TAVYYCAKDP
VH3 1-3 3-30	ISYDGSNKKY	ADSVKGRFTI	SRDNSKNTLY	LQMNSLRAED	TAVYYCAK
Humanized I-3466 HCVR	ISGGGSYTFY	PDSVKGRFTI	SRDNSKNTLY	LQMNSLRAED	TAVYFCTRNO

	101		129
mouse I-3466 HCVR	LLTGMINPLT	TPRAWFTYWG	QGTLVTVSA
BAC02119	RIRLNIG---	DPYYFDYWG	QGTLVTVSS
Humanized I-3466 HCVR	LLTGMINPLT	TPRAWFTYWG	QGTLVTVSS

FIGURE 8

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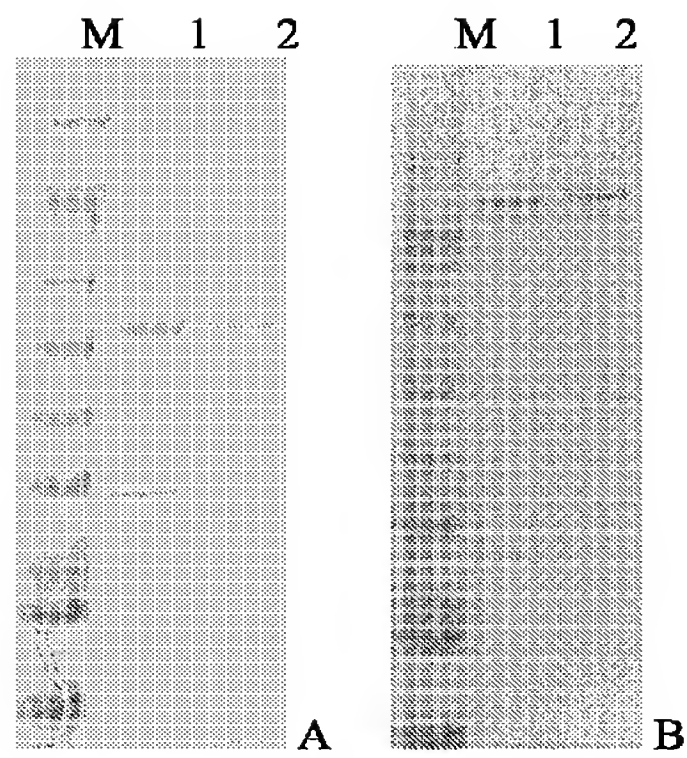


FIGURE 9

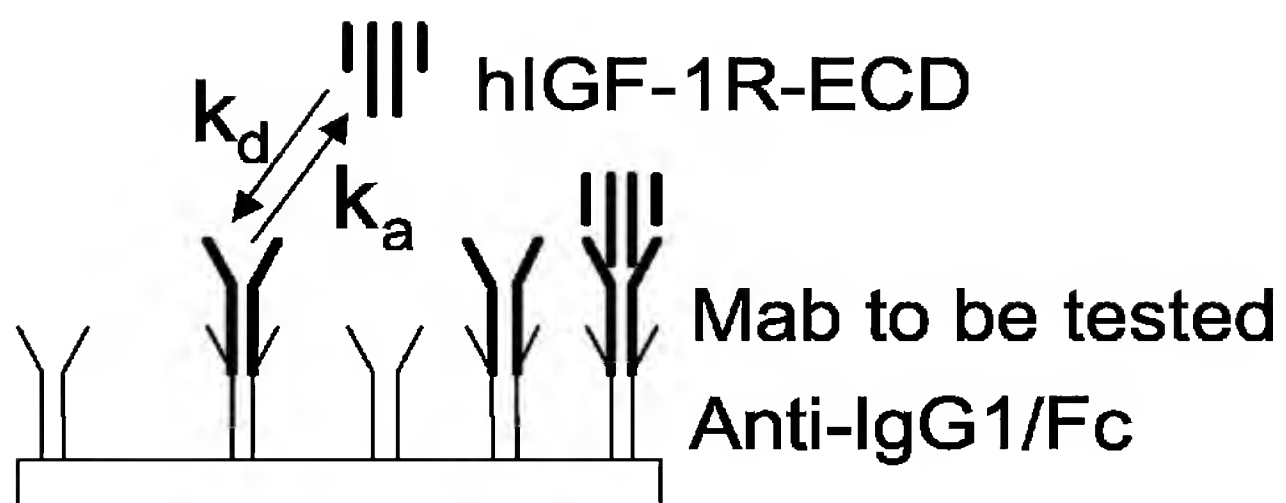


FIGURE 10

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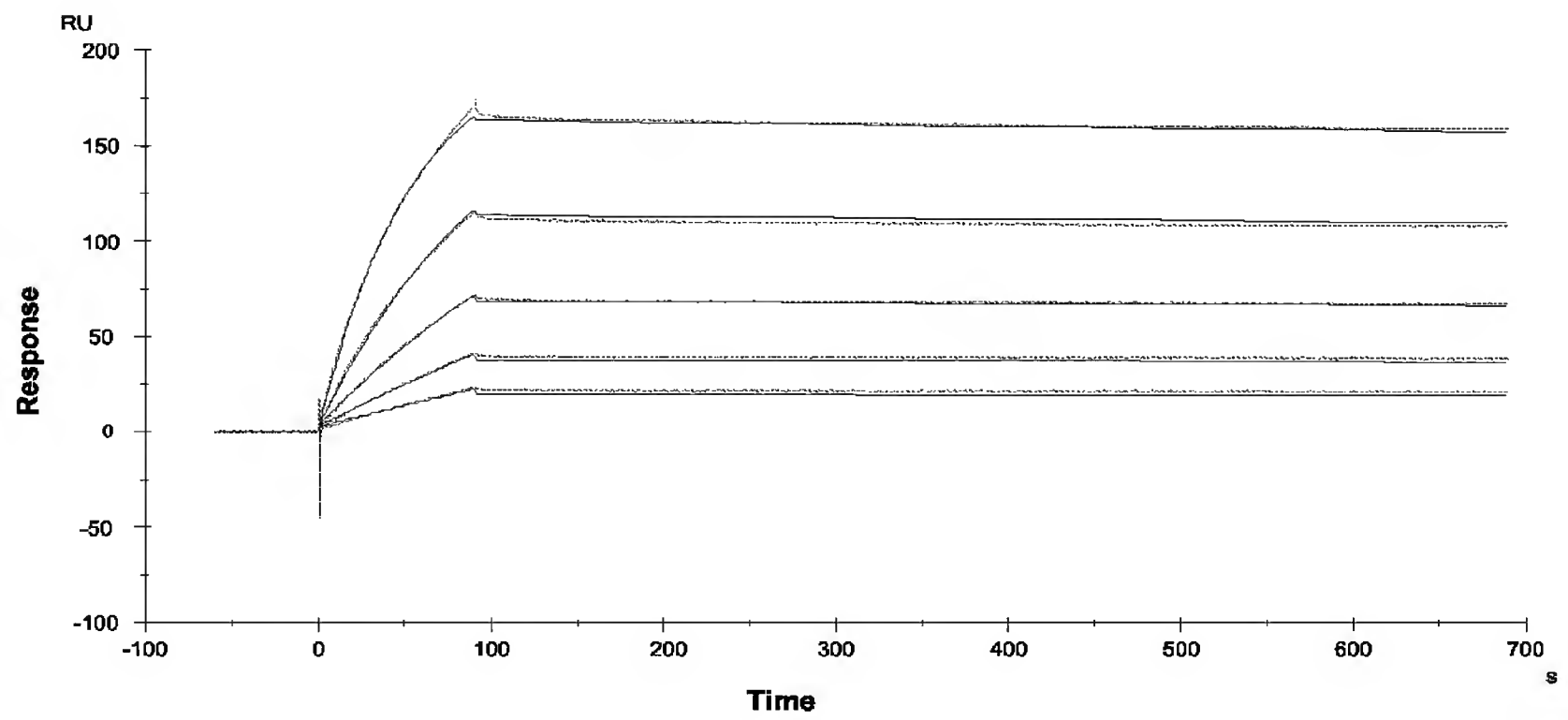


FIGURE 11

PCT

Print Out (Original in Electronic Form)

0-1	Form PCT/RO/134 (SAFE) Indications Relating to Deposited Microorganism(s) or Other Biological Material (PCT Rule 13bis)	
0-1-1	Prepared Using	PCT Online Filing Version 3.5.000.176 MT/FOP 20020701/0.20.4rc.2.7
0-2	International Application No.	
0-3	Applicant's or agent's file reference	349901D23693MIP

1	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
1-1	page	55
1-2	line	4 and 5
1-3	Identification of deposit	
1-3-1	Name of depositary institution	CNCM Collection nationale de cultures de micro-organismes
1-3-2	Address of depositary institution	Institut Pasteur, 28, rue du Dr Roux, 75724 Paris Cedex 15, France
1-3-3	Date of deposit	23 June 2005 (23.06.2005)
1-3-4	Accession Number	CNCM I-3466
1-5	Designated States for Which Indications are Made	all designations

FOR RECEIVING OFFICE USE ONLY

0-4	This form was received with the international application: (yes or no)	
0-4-1	Authorized officer	

FOR INTERNATIONAL BUREAU USE ONLY

0-5	This form was received by the international Bureau on:	
0-5-1	Authorized officer	

FORMULE INTERNATIONALE

DESTINATAIRE :

Christian Bailly
Institut de Recherche Pierre Fabre
Centre d'Immunologique Pierre Fabre
5, avenue Napoléon III - BP 60497
74164 Saint-Julien en Genevois

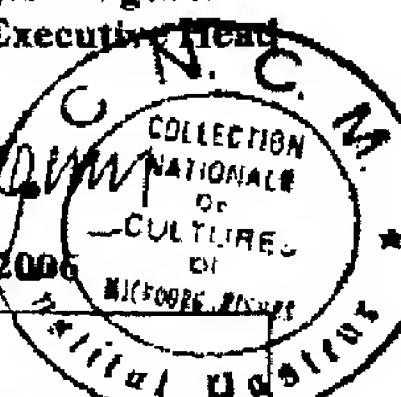
NOM ET ADRESSE
DU DÉPOSANT

RECEVÉ EN CAS DE DÉPÔT INITIAL,
délivré en vertu de la règle 7.1 par
l'AUTORITÉ DE DÉPÔT INTERNATIONALE
identifiée au bas de cette page

Copy of the Original Receipt
certified by

Dr Georges Wagener
CNCM, Executive Head

June 26, 2006



I. IDENTIFICATION DU MICRO-ORGANISME	
Référence d'identification donnée par le DÉPOSANT : 13 G5	Numéro d'ordre attribué par l'AUTORITÉ DE DÉPÔT INTERNATIONALE : CNCM I-3466
II. DESCRIPTION SCIENTIFIQUE ET/OU DÉSIGNATION TAXONOMIQUE PROPOSÉE	
Le micro-organisme identifié sous chiffre I était accompagné :	
<input checked="" type="checkbox"/> d'une description scientifique	
<input checked="" type="checkbox"/> d'une désignation taxonomique proposée	
(Cocher ce qui convient)	
III. RÉCEPTION ET ACCEPTATION	
La présente autorité de dépôt internationale accepte le micro-organisme identifié sous chiffre I, qu'elle a reçu le 23 juin 2005 (date du dépôt initial)	
IV. RÉCEPTION D'UNE REQUÊTE EN CONVERSION	
La présente autorité de dépôt internationale a reçu le micro-organisme identifié sous chiffre I le (date du dépôt initial) et a reçu une requête en conversion du dépôt initial en dépôt conforme au Traité de Budapest le (date de réception de la requête en conversion)	
V. AUTORITÉ DE DÉPÔT INTERNATIONALE	
Nom : COLLECTION NATIONALE DE CULTURES DE MICROORGANISMES (CNCM) Adresse : Institut Pasteur 25, rue du Docteur Roux F-75724 Paris Cedex 15 (France)	Signature(s) de la (des) personne(s) compétente(s) pour représenter l'autorité de dépôt internationale ou de la (des) personne(s) autorisée(s) : Georges Wagener Paris, le 09 septembre 2005

1 En cas d'application de la règle 6.4.d), la date est la date à laquelle le statut
d'autorité de dépôt internationale a été acquis.

**TRAITÉ DE BUDAPEST SUR LA DÉCLARATION
INTERNATIONALE DU DÉPÔT DES MICRO-ORGANISMES
AUX FINS DE LA PROCÉDURE EN MATIÈRE DE BREVETS**

FORMULE INTERNATIONALE

DESTINATAIRE :

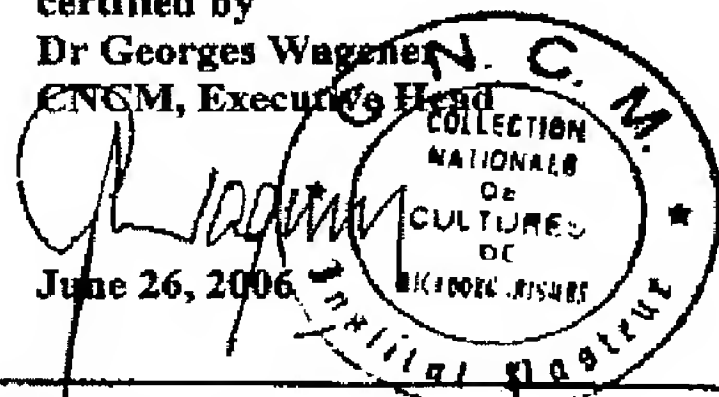
Madame Charlotte BEAU-LARYOR
Madame Liliane GOETSCH
Monsieur Christian BAILLY

INSTITUT DE RECHERCHE PIERRE FABRE
CENTRE D'IMMUNOLOGIE PIERRE FABRE
5, avenue NAPOLEON III - BP 60497
74164 SAINT-JULIEN EN GENEVOIS CEDEX
NOM ET ADRESSE DE LA PARTIE
À LAQUELLE LA DÉCLARATION SUR LA
VIABILITÉ EST DÉLIVRÉE

DÉCLARATION SUR LA VIABILITÉ,
délivrée en vertu de la règle 10.2 par
l'AUTORITÉ DE DÉPÔT INTERNATIONALE
Identifiée à la

Copy of the Last Viability Statement
certified by

Dr Georges Wagener
CNCM, Executive Head



June 26, 2006

I. DEPOSANT	II. IDENTIFICATION DU MICRO-ORGANISME
<p>Nom : Christian Bailly Institut de Recherche Pierre Fabre Centre d'immunologique Pierre Fabre</p> <p>Adresse : 5, avenue Napoléon III - BP 60497 74164 Saint-Julien en Genevois</p>	<p>numéro d'ordre attribué par l'AUTORITÉ DE DÉPÔT INTERNATIONALE : CNCM I-3466</p> <p>date du dépôt ou du transfert : 23 juin 2005</p>
III. DÉCLARATION SUR LA VIABILITÉ	
<p>La viabilité du micro-organisme identifié sous le chiffre III a été contrôlée le 23 juin 2005. A cette date, le micro-organisme</p> <p><input checked="" type="checkbox"/> était viable</p> <p><input type="checkbox"/> n'était plus viable</p>	

- Indiquer la date du dépôt initial ou, si un nouveau dépôt ou un transfert ont été effectués, la plus récente des dates pertinentes (date du nouveau dépôt ou date du transfert).
- Dans les cas visés à la règle 10.2(a)(1) et (11), mentionner le contrôle de viabilité le plus récent.
- Cocher la case qui convient.

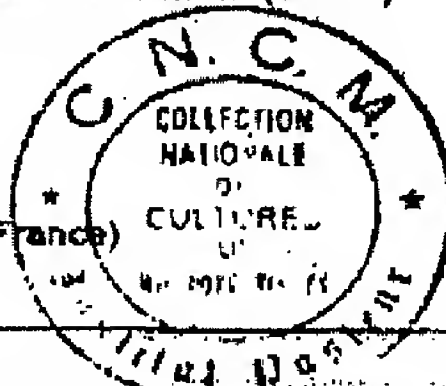
IV. CONDITIONS DANS LESQUELLES LE CONTRÔLE DE VÉRIFICATION A ÉTÉ RÉALISÉ

V. AUTORITÉ DE DÉPÔT INTERNATIONALE

Nom :

COLLECTION NATIONALE
DE CULTURES DE MICROORGANISMES (CNCM)

Adresse :

Institut Pasteur
25, rue du Docteur Roux
F-75724 Paris Cedex 15 (France)Signature(s) de la (des) personne(s)
compétente(s) pour représenter l'autorité
de dépôt internationale ou de la (des)
autorité(s) accrédité(s) :

Georges Wagener

Simona OZDEN
Conseiller Scientifique de la CNCM
pour les virus et cultures cellulaires

Date : Paris, le 09 septembre 2005

À remplir si cette information a été demandée et si les résultats du contrôle étaient négatifs.